

Année 2021-2022

Thèse

Pour le

DOCTORAT EN MEDECINE

Diplôme d'État
par

Elom Annette Amie TAY

Née le 16 novembre 1996 à Lomé (TOGO) (99)

Profil métabolomique et réponse thérapeutique à l'adalimumab au cours des rhumatismes inflammatoires

Présentée et soutenue publiquement le **20 juin 2022** devant un jury composé de :

Président du Jury :

Professeur Philippe GOUPILLE, Rhumatologie, Faculté de Médecine – Tours

Membres du Jury :

Professeur Jérémie SELLAM, Rhumatologie, Faculté de Sorbonne Université – Paris

Professeur Patrick EMOND, Imagerie Moléculaire & Métabolomique, Faculté de Pharmacie – Tours

Professeur Denis MULLEMAN, Rhumatologie, Faculté de Médecine – Tours

Professeur Hélène BLASCO, Biochimie et Biologie Moléculaire, Faculté de Médecine - Tours

Directeurs de thèse : Professeur Denis MULLEMAN & Professeur Hélène BLASCO

UNIVERSITE DE TOURS
FACULTE DE MEDECINE DE TOURS

DOYEN
Pr Patrice DIOT

VICE-DOYEN
Pr Henri MARRET

ASSESSEURS

Pr Denis ANGOULVANT, *Pédagogie*
Pr Mathias BUCHLER, *Relations internationales*
Pr Theodora BEJAN-ANGOULVANT, *Moyens - relations avec l'Université*
Pr Clarisse DIBAO-DINA, *Médecine générale*
Pr François MAILLOT, *Formation Médicale Continue*
Pr Patrick VOURC'H, *Recherche*

RESPONSABLE ADMINISTRATIVE

Mme Fanny BOBLETER

DOYENS HONORAIRES

Pr Emile ARON (†) - 1962-1966
Directeur de l'Ecole de Médecine - 1947-1962
Pr Georges DESBUQUOIS (†) - 1966-1972
Pr André GOUAZE (†) - 1972-1994
Pr Jean-Claude ROLLAND - 1994-2004
Pr Dominique PERROTIN - 2004-2014

PROFESSEURS EMERITES

Pr Daniel ALISON
Pr Gilles BODY
Pr Jacques CHANDENIER
Pr Philippe COLOMBAT
Pr Etienne DANQUECHIN-DORVAL
Pr Pascal DUMONT
Pr Dominique GOGA
Pr Gérard LORETTE
Pr Dominique PERROTIN
Pr Roland QUENTIN

PROFESSEURS HONORAIRES

P. ANTHONIOZ - P. ARBEILLE - A. AUDURIER - A. AUTRET - P. BAGROS - P. BARDOS - C. BARTHELEMY - J.L. BAULIEU - C. BERGER - JC. BESNARD - P. BEUTTER - C. BONNARD - P. BONNET - P. BOUGNOUX - P. BURDIN - L. CASTELLANI - A. CHANTEPIE - B. CHARBONNIER - P. CHOUTET - T. CONSTANS - P. COSNAY - C. COUET - L. DE LA LANDE DE CALAN - J.P. FAUCHIER - F. FETISOF - J. FUSCIARDI - P. GAILLARD - G. GINIES - A. GOUDEAU - J.L. GUILMOT - O. HAILLOT - N. HUTEN - M. JAN - J.P. LAMAGNERE - F. LAMISSE - Y. LANSON - O. LE FLOCH - Y. LEBRANCHU - E. LECA - P. LECOMTE - AM. LEHR-DRYLEWICZ - E. LEMARIE - G. LEROY - M. MARCHAND - C. MAURAGE - C. MERCIER - J. MOLINE - C. MORAIN - J.P. MUH - J. MURAT - H. NIVET - L. POURCELOT - P. RAYNAUD - D. RICHARD-LENOBLE - A. ROBIER - J.C. ROLLAND - D. ROYERE - A. SAINDELLE - E. SALIBA - J.J. SANTINI - D. SAUVAGE - D. SIRINELLI - J. WEILL

PROFESSEURS DES UNIVERSITES - PRATICIENS HOSPITALIERS

ANDRES Christian	Biochimie et biologie moléculaire
ANGOULVANT Denis	Cardiologie
APETOH Lionel.....	Immunologie
AUPART Michel	Chirurgie thoracique et cardiovasculaire
BABUTY Dominique	Cardiologie
BAKHOS David	Oto-rhino-laryngologie
BALLON Nicolas	Psychiatrie ; addictologie
BARILLOT Isabelle	Cancérologie ; radiothérapie
BARON Christophe	Immunologie
BEJAN-ANGOULVANT Théodora	Pharmacologie clinique
BERHOUET Julien	Chirurgie orthopédique et traumatologique
BERNARD Anne	Cardiologie
BERNARD Louis	Maladies infectieuses et maladies tropicales
BLANCHARD-LAUMONNIER Emmanuelle	Biologie cellulaire
BLASCO Hélène	Biochimie et biologie moléculaire
BONNET-BRILHAULT Frédérique.....	Physiologie
BOURGUIGNON Thierry	Chirurgie thoracique et cardiovasculaire
BRILHAULT Jean.....	Chirurgie orthopédique et traumatologique
BRUNEREAU Laurent.....	Radiologie et imagerie médicale
BRUYERE Franck	Urologie
BUCHLER Matthias	Néphrologie
CALAIS Gilles	Cancérologie, radiothérapie
CAMUS Vincent	Psychiatrie d'adultes
CORCIA Philippe	Neurologie
COTTIER Jean-Philippe	Radiologie et imagerie médicale
DEQUIN Pierre-François.....	Thérapeutique
DESOUBEAUX Guillaume	Parasitologie et mycologie
DESTRIEUX Christophe	Anatomie
DIOT Patrice	Pneumologie
DU BOUEXIC de PINIEUX Gonzague	Anatomie & cytologie pathologiques
DUCLUZEAU Pierre-Henri.....	Endocrinologie, diabétologie, et nutrition
EL HAGE Wissam	Psychiatrie adultes
EHRMANN Stephan	Médecine intensive - réanimation
FAUCHIER Laurent	Cardiologie
FAVARD Luc	Chirurgie orthopédique et traumatologique
FOUGERE Bertrand.....	Gériatrie
FOUQUET Bernard	Médecine physique et de réadaptation
FRANCOIS Patrick	Neurochirurgie
FROMONT-HANKARD Gaëlle	Anatomie & cytologie pathologiques
GATAULT Philippe.....	Néphrologie
GAUDY-GRAFFIN Catherine	Bactériologie-virologie, hygiène hospitalière
GOUPILLE Philippe	Rhumatologie
GRUEL Yves	Hématologie, transfusion
GUERIF Fabrice	Biologie et médecine du développement et de la reproduction
GUILLON Antoine.....	Médecine intensive - réanimation
GUYETANT Serge	Anatomie et cytologie pathologiques
GYAN Emmanuel	Hématologie, transfusion
HALIMI Jean-Michel.....	Thérapeutique
HANKARD Régis.....	Pédiatrie
HERAULT Olivier.....	Hématologie, transfusion
HERBRETEAU Denis	Radiologie et imagerie médicale
HOURIOUX Christophe.....	Biologie cellulaire
IVANES Fabrice	Physiologie
LABARTHE François	Pédiatrie
LAFFON Marc	Anesthésiologie et réanimation chirurgicale, médecine d'urgence
LARDY Hubert	Chirurgie infantile
LARIBI Saïd	Médecine d'urgence
LARTIGUE Marie-Frédérique	Bactériologie-virologie
LAURE Boris	Chirurgie maxillo-faciale et stomatologie
LECOMTE Thierry	Gastroentérologie, hépatologie

LESCANNE Emmanuel	Oto-rhino-laryngologie
LINASSIER Claude	Cancérologie, radiothérapie
MACHET Laurent	Dermato-vénérérologie
MAILLOT François	Médecine interne
MARCHAND-ADAM Sylvain	Pneumologie
MARRET Henri	Gynécologie-obstétrique
MARUANI Annabel	Dermatologie-vénérérologie
MEREGHETTI Laurent	Bactériologie-virologie ; hygiène hospitalière
MITANCHEZ Delphine	Pédiatrie
MORINIERE Sylvain	Oto-rhino-laryngologie
MOUSSATA Driffa	Gastro-entérologie
MULLEMAN Denis	Rhumatologie
ODENT Thierry	Chirurgie infantile
OUAISSE Mehdi	Chirurgie digestive
OULDAMER Lobna	Gynécologie-obstétrique
PAINTAUD Gilles	Pharmacologie fondamentale, pharmacologie clinique
PATAT Frédéric	Biophysique et médecine nucléaire
PERROTIN Franck	Gynécologie-obstétrique
PISELLA Pierre-Jean	Ophtalmologie
PLANTIER Laurent	Physiologie
REMERAND Francis	Anesthésiologie et réanimation, médecine d'urgence
ROINGEARD Philippe	Biologie cellulaire
ROSSET Philippe	Chirurgie orthopédique et traumatologique
RUSCH Emmanuel	Epidémiologie, économie de la santé et prévention
SAINT-MARTIN Pauline	Médecine légale et droit de la santé
SALAME Ephrem	Chirurgie digestive
SAMIMI Mahtab	Dermatologie-vénérérologie
SANTIAGO-RIBEIRO Maria	Biophysique et médecine nucléaire
THOMAS-CASTELNAU Pierre	Pédiatrie
TOUTAIN Annick	Génétique
VAILLANT Loïc	Dermato-vénérérologie
VELUT Stéphane	Anatomie
VOURC'H Patrick	Biochimie et biologie moléculaire
WATIER Hervé	Immunologie
ZEMMOURA Ilyess	Neurochirurgie

PROFESSEUR DES UNIVERSITES DE MEDECINE GENERALE

DIBAO-DINA Clarisse LEBEAU Jean-Pierre

PROFESSEURS ASSOCIES

MALLET Donatien	Soins palliatifs
POTIER Alain	Médecine Générale
ROBERT Jean	Médecine Générale

PROFESSEUR CERTIFIE DU 2ND DEGRE

MC CARTHY Catherine

Anglais

MAITRES DE CONFERENCES DES UNIVERSITES - PRATICIENS HOSPITALIERS

AUDEMARD-VERGER Alexandra	Médecine interne
BARBIER Louise	Chirurgie digestive
BINET Aurélien	Chirurgie infantile
BISSON Arnaud	Cardiologie (CHRO)
BRUNAUT Paul	Psychiatrie d'adultes, addictologie
CAILLE Agnès	Biostat., informatique médical et technologies

de communication	
CARVAJAL-ALLEGRIA Guillermo	Rhumatologie (au 01/10/2021)
CLEMENTY Nicolas	Cardiologie
DENIS Frédéric	Odontologie
DOMELIER Anne-Sophie	Bactériologie-virologie, hygiène hospitalière
DUFOUR Diane	Biophysique et médecine nucléaire
ELKRIEF Laure	Hépatologie - gastroentérologie
FAVRAIS Géraldine	Pédiatrie
FOUQUET-BERGEMER Anne-Marie	Anatomie et cytologie pathologiques
GOUILLEUX Valérie	Immunologie
GUILLON-GRAMMATICO Leslie	Epidémiologie, économie de la santé et prévention
HOARAU Cyrille	Immunologie
LE GUELLEC Chantal	Pharmacologie fondamentale, pharmacologie clinique
LEFORT Bruno	Pédiatrie
LEGRAS Antoine	Chirurgie thoracique
LEMAIGNEN Adrien	Maladies infectieuses
MACHET Marie-Christine	Anatomie et cytologie pathologiques
MOREL Baptiste	Radiologie pédiatrique
PARE Arnaud	Chirurgie maxillo-faciale et stomatologie
PIVER Éric	Biochimie et biologie moléculaire
REROLLE Camille	Médecine légale
ROUMY Jérôme	Biophysique et médecine nucléaire
SAUTENET Bénédicte	Thérapeutique
STANDLEY-MIQUELESTORENA Elodie	Anatomie et cytologie pathologiques
STEFIC Karl	Bactériologie
TERNANT David	Pharmacologie fondamentale, pharmacologie clinique
VUILLAUME-WINTER Marie-Laure	Génétique

MAITRES DE CONFERENCES DES UNIVERSITES

AGUILLON-HERNANDEZ Nadia	Neurosciences
NICOGLOU Antonine	Philosophie - histoire des sciences et des techniques
PATIENT Romuald	Biologie cellulaire
RENOUX-JACQUET Cécile	Médecine Générale

MAITRES DE CONFERENCES ASSOCIES

BARBEAU Ludivine	Médecine Générale
ETTORI-AJASSE Isabelle	Médecine Générale
PAUTRAT Maxime	Médecine Générale
RUIZ Christophe	Médecine Générale
SAMKO Boris	Médecine Générale

CHERCHEURS INSERM - CNRS - INRAE

BECKER Jérôme	Chargé de Recherche Inserm - UMR Inserm 1253
BOUAKAZ Ayache	Directeur de Recherche Inserm - UMR Inserm 1253
BRIARD Benoit	Chargé de Recherche Inserm - UMR Inserm 1100
CHALON Sylvie	Directeur de Recherche Inserm - UMR Inserm 1253
DE ROCQUIIGNY Hugues	Chargé de Recherche Inserm - UMR Inserm 1259
ESCOFFRE Jean-Michel	Chargé de Recherche Inserm - UMR Inserm 1253
GILOT Philippe	Chargé de Recherche Inrae - UMR Inrae 1282
GOUILLEUX Fabrice	Directeur de Recherche CNRS - EA 7501 - ERL CNRS 7001
GOMOT Marie	Chargée de Recherche Inserm - UMR Inserm 1253
HEUZE-VOURCH Nathalie	Directrice de Recherche Inserm - UMR Inserm 1100
KORKMAZ Brice	Chargé de Recherche Inserm - UMR Inserm 1100
LATINUS Marianne	Chargée de Recherche Inserm - UMR Inserm 1253
LAUMONNIER Frédéric	Chargé de Recherche Inserm - UMR Inserm 1253
LE MERREUR Julie	Directrice de Recherche CNRS - UMR Inserm 1253
MAMMANO Fabrizio	Directeur de Recherche Inserm - UMR Inserm 1259
MEUNIER Jean-Christophe	Chargé de Recherche Inserm - UMR Inserm 1259
PAGET Christophe	Chargé de Recherche Inserm - UMR Inserm 1100

RAOUL William	Chargé de Recherche Inserm - UMR CNRS 1069
SI TAHAR Mustapha	Directeur de Recherche Inserm - UMR Inserm 1100
SUREAU Camille	Directrice de Recherche émérite CNRS - UMR Inserm 1259
WARDAK Claire	Chargée de Recherche Inserm - UMR Inserm 1253

CHARGES D'ENSEIGNEMENT

Pour l'Ecole d'Orthophonie

DELORE Claire	Orthophoniste
GOUIN Jean-Marie	Praticien Hospitalier

Pour l'Ecole d'Orthoptie

BOULNOIS Sandrine	Orthoptiste
SALAME Najwa	Orthoptiste

Pour l'Ethique Médicale

BIRMELE Béatrice	Praticien Hospitalier
------------------------	-----------------------

SERMENT D'HIPPOCRATE

En présence des Maîtres de cette Faculté,
de mes chers condisciples et selon la
tradition d'Hippocrate, je promets et je
jure d'être fidèle aux lois de l'honneur et
de la probité dans l'exercice de la
Médecine.

Je donnerai mes soins gratuits à l'indigent,
et n'exigerai jamais un salaire au-dessus de mon travail.

Admis dans l'intérieur des maisons, mes yeux
ne verront pas ce qui s'y passe, ma langue taira
les secrets qui me seront confiés et mon état ne servira pas à
corrompre les mœurs ni à favoriser le crime.

Respectueux et reconnaissant envers mes Maîtres, je
rendrai à leurs enfants
l'instruction que j'ai reçue de leurs pères.

Que les hommes m'accordent leur estime
si je suis fidèle à mes
promesses. Que je sois
couvert d'opprobre et
méprisé de mes confrères
si j'y manque.

REMERCIEMENTS

Mesdames et messieurs les membres du jury,

A Monsieur le Professeur Philippe GOUPILLE, je vous remercie pour avoir accepté de présider ce jury.

A Monsieur le Professeur Jérémie SELLAM, merci de me faire l'honneur de juger ce travail ainsi que pour la conférence ayant précédé la soutenance de ma thèse.

A Monsieur le Professeur Patrick EMOND, je vous remercie d'avoir accepté de participer au jury de cette thèse et pour la supervision scientifique de ce travail.

A Monsieur le Professeur Denis MULLEMAN, je vous remercie de m'avoir proposé ce travail ainsi que pour votre accompagnement tout au long de la rédaction de cette thèse. Merci pour vos encouragements et pour votre disponibilité.

A Madame le Professeur Hélène BLASCO, je vous remercie de m'avoir initié au domaine de la métabolomique à travers cette thèse. Merci pour vos conseils avisés concernant la rédaction du manuscrit.

Aux équipes médicales et paramédicales des services de Rhumatologie à Tours et à Orléans :

Dr Isabelle GRIFFOUL, merci pour la cheffe de service à l'écoute et proche de son équipe que tu es. Merci pour ta rigueur et pour toutes les notions que j'ai pu acquérir à ton contact dès mon premier semestre.

Dr Delphine CHU MIOW LIN, merci pour ton caractère pédagogue tout en douceur, aussi bien dans le service qu'au volant. Merci pour ta disponibilité, ta gentillesse et ta régulière bonne humeur.

Dr Salou MAMMOU, merci pour tout ce que tu m'as apporté en ce qui concerne les relations humaines en plus des connaissances théoriques nécessaires à notre métier. Merci pour ton implication dans notre apprentissage de l'imagerie par échographie en rhumatologie.

Jessica et Guillermo, merci pour votre proximité et vos astuces pour être des pros en échographie ostéoarticulaire.

Florence et Micheline, merci pour votre bonne humeur, votre côté maternel, vos petites blagues et les pauses goûter aussi bien à 10h qu'à 16h au sein de votre bureau, lieu privilégié pour souffler un peu au cours de nos journées marathon.

Aux infirmières Agnès, Corinne, Sandrine, Christine, Elsa, Chrystelle, Virginie, Valérie F. et Valérie A. merci pour l'accueil que vous faites aux internes et pour votre professionnalisme au quotidien.

A Valérie BASSET, meilleure cadre de service que j'ai pu connaître tout au long de mon parcours d'interne.

Dr Nada IBRAHIM, cheffe de service de rhumatologie à Orléans, merci pour ta joie de vivre à toute épreuve et pour ton enseignement au cours des visites.

A mes co-internes et autres chefs, aussi bien en rhumatologie qu'en dehors :

Marie, Naomi, Romane, Johan, Heidi, Camille, Cléa, Amaury, Thibault, Sara et Léa ; merci pour tous les moments partagés ensemble en stage. Je vous souhaite à tous une bonne continuation et de belles carrières.

Alexis, Simon et Kim, merci de m'avoir donné un aperçu de ce que peut être un futur médecin interniste et pour l'ambiance bonne enfant en stage. Alexandre, merci pour l'entraide et les fous rires.

Emmanuelle, Sophie, Alice, Juliette, Fred, Laura, Jordane, Margueritte, Claire LH., Claire D., Anne et Antoine, merci de m'avoir fait découvrir la dermatologie, cette belle spécialité qui ne s'arrête pas qu'aux « boutons ».

A l'équipe du laboratoire de métabolomique et d'analyse chimique de Tours

Merci à M. Antoine LEFEVRE, ingénieur de recherche, à qui l'on doit les analyses des échantillons de l'étude, sans qui ce travail n'aurait pas été possible.

A mes amies :

Angela, Audrey A., Sonia, Murielle, merci pour votre soutien indéfectible depuis toutes ces années malgré la distance.

Linh, Nathalie, Edwige L. et Audrey J., ça y est, on voit le bout. Merci pour les partages d'expérience et pour le soutien mutuel au cours de ces années d'études de médecine, pas toujours faciles à vivre mais grâce auxquelles on fait de belles rencontres.

Enfin, à ma famille :

Ma mère Anne AFETSE, épouse TAY ; ma sœur Akpe, mes tantes (Nodi Enyovi, Jacky, Edoh), mes oncles (Blaise « frère » et Jean AFETSE), merci pour votre soutien inégalé depuis mon enfance quasiment, vous, aux yeux de qui j'étais déjà « docteur » avant même le début de mes études de médecine.

A ma grand-mère Hélène KLUGA épouse AFETSE, qui n'est plus de ce monde mais qui a été ma fidèle compagne pendant plusieurs années à la maison.

A mon mari, Lionel, merci pour ta patience, merci de me soutenir au quotidien, d'essayer de me faire voir les choses autrement et de me faire « vivre » autant que possible malgré la quantité de choses que je pense devoir gérer au quotidien.

A mes cousins Diane et Dorian, à mon beau-frère Claudel ; merci pour la figure d'ainés que vous incarnez dans ma vie, pour votre présence et votre soutien aussi bien dans mes projets individuels qu'à deux avec Lionel.

Pour finir, à feu Docteur TAY Komi, mon père, qui nous a quitté il y a deux ans, celui qui m'a initié à cette belle discipline qu'est la médecine, sans qui je ne me serais probablement pas engagée dans ces études. Merci pour nos échanges presque depuis l'école primaire, merci pour la motivation que tu m'apportais au quotidien, pour cette capacité que tu avais à me pousser vers le haut. « Docteur TAY Junior » est bientôt dans la place.

RESUME

Objectifs : Analyser les changements métabolomiques 4 semaines après l'initiation de l'adalimumab, un anti-TNF chez les patients atteints de spondylarthrite axiale (SpA) et de polyarthrite rhumatoïde (PR) et étudier la relation avec la réponse clinique à 12 et 26 semaines.

Méthodes : Nous avons effectué des analyses en post-hoc du sérum de patients atteints de SpA de l'étude COMARIS (NCT01895764) et de patients atteints de PR de l'étude AFORA (NCT01382160) par chromatographie liquide couplée à la spectrométrie de masse à haute résolution (LC/HRMS) à l'inclusion (S0) et 4 semaines (S4) après l'initiation du traitement par adalimumab. Les réponses cliniques ont été évaluées à 12 et 26 semaines. Les variations des concentrations de métabolites entre S0 et S4 ont été comparées entre les répondeurs et les non-répondeurs en analyse univariée, multivariée non supervisée en composantes principales (PCA) puis multivariée supervisée par des analyses discriminantes partielles des moindres carrés (PLS-DA), à l'aide du logiciel METABOANALYST. Les variables d'intérêt, définies comme celles ayant un score VIP (Variable Influence on Projection) supérieur ou égal à 2 ont été sélectionnées. Enfin, nous avons étudié les différentes voies métaboliques dans lesquelles ces variables d'intérêt étaient impliquées.

Résultats : Soixante-quatorze patients ont été inclus dans la cohorte COMARIS, dont 43 répondeurs et 31 non-répondeurs. Soixante-trois patients ont été inclus dans la cohorte AFORA, dont 52 répondeurs et 11 non-répondeurs. En analyse univariée, la leucine, l'hypoxanthine et la n-acétyl-l-alanine étaient abaissées chez les répondeurs atteints de SpA. La carnosine, le cortisol et la purine étaient abaissés chez les répondeurs atteints de PR, tandis que la l-méthionine et la 5-oxo-l-proline étaient augmentées chez ces sujets. Nous n'avons pas été en mesure de différencier les patients de chaque cohorte en fonction de leur réponse en utilisant la PCA et la PLS-DA. Cependant, chez les patients atteints de SpA, cinq métabolites avec un score VIP >2 de l'analyse PLS-DA, y compris le PAF-C16, un facteur d'activation plaquettaire, et le LysoPC (18:0), un composant des membranes cellulaires, semblaient pertinents. Diverses voies métaboliques ont été identifiées dans les deux cohortes : métabolisme des glycérophospholipides ; métabolisme de l'arginine et de la proline ; métabolisme de lalanine, de l'aspartate et du glutamate ; biosynthèse de la phénylalanine, de la tyrosine et du tryptophane; métabolisme de la cystéine et de la méthionine; biosynthèse du pantothéate et CoA, et biosynthèse de l'aminoacyl-ARNt.

Conclusion : Chez les patients atteints de PR et de SpA, l'adalimumab induit des changements précoces dans le métabolome impliquant des voies telles que le métabolisme des acides aminés essentiels et non essentiels et le stress oxydatif. Ce résultat pourrait guider les investigations futures afin de trouver des marqueurs prédictifs de la réponse thérapeutique aux anti-TNF dans les rhumatismes inflammatoires chroniques.

Mots-clés : métabolomique ; spondylarthrite axiale ; polyarthrite rhumatoïde ; adalimumab ; prédiction de la réponse ; LC/HRMS ; acides aminés ; stress oxydatif.

ABSTRACT

Objectives: To analyse metabolomic changes 4 weeks after initiation of adalimumab, a TNF inhibitor in patients with axial spondyloarthritis (SpA) and Rheumatoid Arthritis (RA) and to study the relationship with clinical response at 12 and 26 weeks.

Methods: We performed post-hoc analyses of serum from patients with SpA from the COMARIS study (NCT01895764) and from patients with RA from the AFORA study (NCT01382160) by liquid chromatography coupled to high-resolution mass spectrometry (LC/HRMS) at inclusion (W0) and 4 weeks (W4) after initiation of adalimumab therapy. Clinical responses were assessed at 12 and 26 weeks. The variations in metabolites concentrations between W0 and W4 were compared between responders and non-responders in univariate, multivariate unsupervised principal component analysis (PCA) and supervised partial least squares discriminant analysis (PLS-DA), using METABOANALYST software. The variables of interest, defined as those with a VIP (Variable Influence on Projection) score greater than or equal to 2 were selected. Finally, we studied the different metabolic pathways in which these variables of interest were involved.

Results: Seventy-four patients were included in the COMARIS cohort, including 43 responders and 31 non-responders. Sixty-three patients were included in the AFORA cohort, including 52 responders and 11 non-responders. In univariate analysis, leucine, hypoxanthine, and n-acetyl-l-alanine were lowered in SpA responders. Carnosine, cortisol, and purine were lowered in RA responders, whereas l-methionine and 5-oxo-l-proline were increased in these subjects. We were unable to differentiate the patients from each cohort according to their response using both PCA and PLS-DA. However, in SpA patients, five metabolites with a VIP-score >2 from the PLS-DA analysis including PAF-C16, a platelet activating factor, and LysoPC(18:0), a component of cell membranes, appeared relevant. Various metabolic pathways were identified in both studies: glycerophospholipid metabolism; arginine and proline metabolism; metabolism of alanine, aspartate, and glutamate; biosynthesis of phenylalanine, tyrosine, and tryptophan; cysteine and methionine metabolism; biosynthesis of pantothenate and CoA, and biosynthesis of aminoacyl-tRNA.

Conclusion: In patients with RA and SpA, adalimumab induced early changes in the metabolome involving pathways such as essential and non-essential amino acid metabolism and oxidative stress. This finding may guide future investigations to find predictive markers of therapeutic response to TNF inhibitors in chronic inflammatory rheumatic diseases.

Keywords: metabolomics; spondylarthritis; rheumatoid arthritis; adalimumab; response prediction; LC/HRMS; amino acids; oxidative stress.

LISTE DES ABBREVIATIONS

ACR	American College of Rheumatology
AFORA in Rheumatoid Arthritis	Serum Concentration of Adalimumab as a Predictive Factor of clinical Outcomes
ARNt	Acide Ribonucléique de transfert
ASDAS	Ankylosing Spondylitis Disease Activity Score
ATRA	all-trans-retinoic acid
bDMARDs	biologic Disease Modifying Anti-Rheumatic Drug
CoA	Coenzyme A
COMARIS	Effect of the Combination of Methotrexate and Adalimumab on Reduction of Immunization in Ankylosing Spondylitis
CRP	C-reactive protein
csDMARDs	conventional synthetic Disease Modifying Anti-Rheumatic Drug
DAS28	Disease Activity Score on 28 joints
ESR	Erythrocyte Sedimentation Rate
EULAR	European Alliance of Associations for Rheumatology
HLA-B27	Human Leukocyte Antigen B27
HUGO	Hôpitaux Universitaires du Grand Ouest
IPP	Interphalangeal proximal
LC/HRMS	Liquid chromatography coupled to high-resolution mass spectrometry
LPC (18:0)	Lysophosphatidylcholine 18:0
MCP	Metacarpophalangeal
MRI	Magnetic Resonance Imaging
PAF	Platelet Activating Factor
PCA	Principal Component Analysis
PLS-DA	Partial Least Squares Discriminant Analysis
RA	Rheumatoid Arthritis
RF	Rheumatoid Factor
S0, S4, S12, S26	Semaine 0, semaine 4, semaine 12, semaine 26
SpA	Spondyloarthritis/Spondylarthrite axiale
TNF	Tumor Necrosis Factor
tRNA	transfert Ribonucleic Acid
tsDMARDs	targeted synthetic Disease Modifying Anti-Rheumatic Drugs
VIP-score	Variable Influence on Projection score
VS	Vitesse de sédimentation
W0, W4, W12, W26	Week 0, week 4, week 12, week 26

TABLE DES MATIERES

Résumé général du travail de thèse	16
1. Introduction.....	16
1.1. Spondyloarthrite et Polyarthrite rhumatoïde	16
1.2. Thérapeutiques actuellement disponibles	17
2. Le sujet de réflexion	17
3. Les projets MetAFORA et MetCOMARIS	17
Metabolomic profile and therapeutic response to adalimumab in inflammatory rheumatic diseases.....	19
Abstract.....	19
Introduction	20
Material and methods	20
1. Study plan	20
2. Detailed study methodology	21
a. Patients.....	21
b. Clinical data	21
c. Blood samples and biological data	21
d. Evaluation of the therapeutic response	22
e. Metabolomic data	22
3. Statistical analysis.....	23
Results	23
1. Population characteristics	23
2. Metabolomic analyses	24
2.1. COMARIS	24
a. Main analysis: based on therapeutic response at 12 weeks	24
b. Secondary analysis: based on therapeutic response at 26 weeks.....	26
c. Common metabolic pathways	27
2.2. AFORA.....	27
a. Main analysis: based on therapeutic response at 12 weeks	27
b. Secondary analysis: based on therapeutic response at 26 weeks.....	29
c. Common metabolic pathways	30
2.3. Comparison of the two cohorts.....	30
Discussion	31
Conclusion.....	34
References	35
Annexes	41

TABLE DES ILLUSTRATIONS

Figure 1. Flow chart of the COMARIS and AFORA cohorts.....	24
Table 1. Baseline characteristics by adalimumab response at 12 Weeks.....	24
Figure 2. COMARIS. PLS-DA separation of different patient groups according to metabolome change between W0 and W4 based on clinical response at 12 weeks.....	25
Figure 3. COMARIS. Score-plot of metabolites of interest of different patient groups according to metabolome change between W0 and W4 as a function of clinical response at 12 weeks..	26
Figure 4. COMARIS. Metabolic pathways involved in the metabolome change between W0 and W4 according to the therapeutic response at 12 weeks.....	26
Figure 5. COMARIS. Venn diagram representing common metabolic pathways in metabolome analyses between W0 and W4 according to therapeutic responses at 12 and 26 weeks	27
Figure 6. AFORA. PLS-DA separation of different patient groups according to metabolome change between W0 and W4 based on clinical response at 12 weeks.	28
Figure 7. AFORA. Score-plot of metabolites of interest of different patient groups according to metabolome change between W0 and W4 according to clinical response at 12 weeks.	29
Figure 8. AFORA. Metabolic pathways involved in the metabolome change between W0 and W4 according to therapeutic response at 12 weeks.....	29
Figure 9. AFORA. Venn diagram representing common metabolic pathways for metabolome analyses between W0 and W4 according to therapeutic responses at 12 and 26 weeks	30
Figure 10. Metabolic pathways common to both cohorts based on metabolome change between W0 and W4.....	31

Résumé général du travail de thèse

1. Introduction

1.1. Spondyloarthrite et Polyarthrite rhumatoïde

Le terme « spondyloarthrite » désigne un groupe de maladies inflammatoires chroniques caractérisées par des points communs sur le plan clinique et génétique. Ces pathologies peuvent toucher aussi bien le squelette axial que les articulations périphériques. La prévalence de l'atteinte axiale (Spondylarthrite Ankylosante) dans la population mondiale varie entre 0,32% et 1,4% avec des particularités régionales et un gradient Nord-Sud (1).

Cliniquement, les patients atteints de spondylarthrite axiale présentent des douleurs de rythme inflammatoire du rachis ainsi que des fesses, parfois à bascule avec une importante raideur matinale. Parmi les manifestations périphériques les plus fréquentes, nous pouvons citer les arthrites et les enthésites retrouvées chez 30 à 50% des patients atteints de spondylarthrite axiale au diagnostic ou ayant présenté ces symptômes dans les années précédant le diagnostic. Enfin, des symptômes extra-articulaires tels que l'uvéite antérieure – atteinte extra-articulaire la plus fréquente – peuvent venir compléter le tableau clinique (1).

Sur le plan pathogénique, de multiples mécanismes sont à noter tels qu'une prédisposition génétique faisant intervenir certains antigènes du complexe majeur d'histocompatibilité, le plus connu étant le gène HLA-B27 (2). Des mécanismes immunitaires sont également à mentionner, notamment ceux faisant intervenir des cytokines pro-inflammatoires telles que le Tumor Necrosis Factor (TNF) et les interleukines 23 et 17, dont les concentrations sont élevées dans le sérum des patients atteints de spondylarthrite (3). Cependant, l'ensemble de la physiopathologie n'est pas totalement connu à ce jour.

La polyarthrite rhumatoïde, quant à elle, touche environ 0,5 à 1% de la population mondiale avec des variations régionales (4). En France, elle est estimée à 0,3% avec là encore des particularités régionales (5). Sur le plan clinique, elle se présente comme une atteinte symétrique des petites et moyennes articulations avec des douleurs de rythme inflammatoire et une importante raideur matinale. Sa physiopathologie n'est pas non plus complètement connue mais elle serait la conséquence d'une interaction entre des facteurs environnementaux tels que le tabac (6,7), le changement dans le microbiote buccal et intestinal (8) et un terrain génétique prédisposé. La combinaison de ces éléments entraînerait un état inflammatoire au niveau des membranes synoviales faisant intervenir à nouveau le TNF (9).

En ce qui concerne l'activité de ces pathologies, celle-ci peut être évaluée par des scores tels que l'ASDAS (Ankylosing Spondylitis Disease Activity Score) développé en 2009 pour la spondyloarthrite (10,11) et le DAS28 (Disease Activity Score 28), développé dans les années 90 pour la polyarthrite rhumatoïde (12) qui sont tous les deux des scores composites tenant compte de l'auto-évaluation de la maladie par le patient associée à celle du clinicien et d'un marqueur biologique de l'inflammation, la Protéine C-réactive (CRP) ou la vitesse de sédimentation (VS). La spondylarthrite est considérée comme inactive pour un score ASDAS inférieur à 1,3 ; modérée pour un score compris entre 1,3 et 2,1 ; active pour un score compris entre 2,1 et 3,5 et étant très active pour un score au-delà de 3,5 (13). La polyarthrite rhumatoïde est considérée comme étant en rémission pour un score DAS28 inférieur à 2,6 ; d'activité faible pour un score compris entre 2,6 et 3,2 ; d'activité modérée pour un score compris entre 3,2 et 5,1 et enfin d'activité élevée pour un score supérieur à 5,1 (14,15). Ainsi, il est possible de définir la réponse thérapeutique de ces deux pathologies en fonction de la variation de leur score d'activité entre l'introduction du traitement et la réévaluation par le clinicien.

1.2. Thérapeutiques actuellement disponibles

La prise en charge de la spondylarthrite axiale et de la polyarthrite rhumatoïde repose sur l'utilisation de traitements symptomatiques tels que les anti-inflammatoires non stéroïdiens, la corticothérapie orale et les traitements de fond désignés aujourd'hui par le terme csDMARDs pour « conventional synthetic Disease Modifying Anti-Rheumatic Drug » (Méthotrexate, Leflunomide, Sulfasalazine notamment) en première intention. Depuis plusieurs années, les traitements de fond se sont étoffés d'alternatives que sont des molécules ciblant des voies d'activation cellulaire ou des marqueurs de l'inflammation appelés bDMARDs pour « biologic Disease Modifying Anti-Rheumatic Drug » ou tsDMARDs pour « targeted synthetic Disease Modifying Anti-Rheumatic Drugs » (**16**). Parmi celles-ci, on distingue :

- bDMARDs :
 - Les anti-TNF : adalimumab ; certolizumab ; etanercept ; golimumab ; infliximab
 - Les anti-interleukine 6 : tocilizumab ; sarilumab
 - Le modulateur de la costimulation des lymphocytes T : abatacept
 - Le rituximab
- tsDMARDs : baricitinib ; tofacitinib ; upadacitinib

Ces molécules peuvent être et sont de plus en plus utilisées en association ou en monothérapie chez les sujets ayant une réponse incomplète ou une intolérance aux traitements de première ligne dans un but de rémission ou de moindre activité de leur pathologie (**17–19**).

2. Le sujet de réflexion

Malgré les nombreuses options thérapeutiques dont le rhumatologue dispose, 30 à 40% des patients atteints de spondylarthrite axiale ou de polyarthrite rhumatoïde ne répondent pas complètement au traitement (**20–22**). Aujourd'hui, en plus de la compréhension des mécanismes physiopathologiques de ces deux rhumatismes inflammatoires chroniques, il devient nécessaire d'identifier des marqueurs prédictifs de la réponse afin de pouvoir proposer une prise en charge thérapeutique la plus optimale possible.

La métabolomique désigne l'exploration de l'ensemble des métabolites des organismes vivants et fait partie du groupe des « OMICS » (génomique, transcriptomique, protéomique, lipidomique et glucidomique), procédés développés ces dernières années ayant permis d'explorer et de comprendre de nombreux processus physiopathologiques sur le plan moléculaire. L'inflammation locale et systémique retrouvée dans les rhumatismes inflammatoires chroniques est à l'origine de modifications dans le métabolisme local et général, pouvant être objectivées par les analyses métabolomiques (**23**). Des profils métaboliques particuliers, pouvant correspondre à des « signatures » de ces pathologies ont ainsi pu être mis en évidence (**24–26**).

Sur le plan thérapeutique, les modifications induites dans le métabolome par les traitements tels que les anti-TNF alpha ont pu être étudiées chez les patients atteints de spondylarthrite d'une part et de polyarthrite rhumatoïde d'autre part, révélant la restauration de l'équilibre de certains métabolites dans le sérum après traitement chez les premiers et la possibilité de différencier les répondeurs et les non répondeurs au traitement selon le profil métabolique urinaire de base chez les seconds par exemple (**27,28**).

3. Les projets MetaFORA et MetCOMARIS

C'est ainsi que sont nés les projets MetaFORA et MetCOMARIS, consistant en l'analyse post-hoc, en utilisant la spectrométrie de masse, d'échantillons de sérum de patients des études AFORA (Serum Concentration of Adalimumab as a Predictive Factor of clinical Outcomes in Rheumatoid Arthritis, NCT01382160) et COMARIS (Effect of the Combination of

Methotrexate and Adalimumab on Reduction of Immunization in Ankylosing Spondylitis, NCT01895764) dans le but d'observer des trajectoires métaboliques selon la réponse à l'adalimumab afin d'identifier des biomarqueurs prédictifs.

Dans le cadre de ces études, des prélèvements sanguins avaient été réalisés avant et après initiation du traitement par adalimumab à intervalles réguliers (4 semaines, 8 semaines, 12 semaines puis 26 semaines) en parallèle de l'évaluation de la réponse clinique au traitement via les scores d'activité. Les résultats de l'étude COMARIS sont publiés et disponibles (29); ceux de l'étude AFORA sont encore en cours d'analyse.

MetAFORA a initialement fait l'objet de la thèse de médecine du Dr Caroline PETIT, soutenue en septembre 2020. Ses analyses avaient permis de mettre en évidence une différence de concentration de certains métabolites avant traitement par adalimumab selon la réponse EULAR et l'hippurate semblait se démarquer parmi les métabolites potentiellement prédictifs de la réponse thérapeutique. Des éléments cliniques et métabolomiques des patients de l'étude COMARIS étant disponibles, nous avons envisagé de faire le même type de travail sur ces données (naissance de l'étude MetCOMARIS), de reprendre celles de l'étude MetAFORA, puis de comparer les résultats entre les deux études.

Ainsi, dès novembre 2021, nous avons commencé par une première phase de bibliographie afin de faire le point sur la littérature disponible dans le domaine de la métabolomique et les pathologies rhumatologiques inflammatoires chroniques. Puis j'ai collecté les caractéristiques cliniques des patients de l'étude COMARIS afin de déterminer leur réponse thérapeutique à l'adalimumab selon la variation du score ASDAS entre l'inclusion et l'évaluation clinique à 12 et 26 semaines. Les données métabolomiques avaient été analysées à l'inclusion, 4 semaines puis 12 semaines par la plateforme PST ASB de l'INSERM 1253, iBrain, et ont été recueillies au mois de décembre 2021.

Le pourcentage de variation des concentrations relatives des métabolites entre l'inclusion et 4 semaines a été déterminé, et une base de données intégrant les données cliniques et métabolomiques a ainsi été constituée en rapportant la différence brute à la valeur de détection de concentration de métabolite la plus faible à l'inclusion. Les données ont été analysées grâce au logiciel METABOANALYST 5.0 afin d'évaluer les profils métabolomiques des sujets répondeurs et non répondeurs au traitement par adalimumab et d'identifier les principales voies métaboliques potentiellement impliquées aussi bien dans la physiopathologie que dans la réponse au traitement.

Dans le but d'assurer une homogénéité de méthodologie entre les deux études, j'ai repris les éléments de MetAFORA selon le même principe à partir du mois de février 2022 : collecte des données cliniques avec identification des répondeurs et non répondeurs selon la variation du score DAS28 entre l'inclusion et l'évaluation clinique à 12 puis 26 semaines en accord avec les recommandations EULAR ; détermination du pourcentage de variation de la concentration des métabolites avec Microsoft Excel entre l'inclusion et 4 semaines puis analyse métabolomique sur METABOANALYST 5.0 dans le même but que celui précédemment cité pour MetCOMARIS.

L'article rapportant les résultats obtenus est présenté au cours des pages suivantes comme travail de thèse. Celui-ci sera soumis à la revue Metabolites (ISSN 2218-1989, IF 4.932), sur invitation le 15 décembre 2021 à participer au numéro spécial intitulé "How Metabolomics Findings Can Drive New Therapeutic Approaches ? » (Guest Editor: Professor Helene Blasco).

Metabolomic profile and therapeutic response to adalimumab in inflammatory rheumatic diseases

Elom Tay^{*1,2}, Caroline Petit^{*1,2}, Patrick Emond^{3,4,5}, Antoine Lefevre^{3,5}, Philippe Goupille¹, Eric Piver⁴, Hélène Blasco^{2#}, Denis Mulleman^{1,2}

¹EA7501 GICC, University of Tours, Tours, France

²Rheumatology Department, CHRU de Tours, Tours, France

³INSERM 1253, iBrain, University of Tours, Tours, France

⁴Biochemistry and Molecular Biology Department, CHRU de Tours, Tours, France

⁵Molecular Imaging and Metabolomics Department, CHRU de Tours, Tours, France

*: contributed equally

#: corresponding author

Abstract

Objectives: To analyse metabolomic changes 4 weeks after initiation of adalimumab, a TNF inhibitor in patients with axial spondyloarthritis (SpA) and Rheumatoid Arthritis (RA) and to study the relationship with clinical response at 12 and 26 weeks.

Methods: We performed post-hoc analyses of serum from patients with SpA from the COMARIS study (NCT01895764) and from patients with RA from the AFORA study (NCT01382160) by liquid chromatography coupled to high-resolution mass spectrometry (LC/HRMS) at inclusion (W0) and 4 weeks (W4) after initiation of adalimumab therapy. Clinical responses were assessed at 12 and 26 weeks. The variations in metabolites concentrations between W0 and W4 were compared between responders and non-responders in univariate, multivariate unsupervised principal component analysis (PCA) and supervised partial least squares discriminant analysis (PLS-DA), using METABOANALYST software. The variables of interest, defined as those with a VIP (Variable Influence on Projection) score greater than or equal to 2 were selected. Finally, we studied the different metabolic pathways in which these variables of interest were involved.

Results: Seventy-four patients were included in the COMARIS cohort, including 43 responders and 31 non-responders. Sixty-three patients were included in the AFORA cohort, including 52 responders and 11 non-responders. In univariate analysis, leucine, hypoxanthine, and n-acetyl-l-alanine were lowered in SpA responders. Carnosine, cortisol, and purine were lowered in RA responders, whereas l-methionine and 5-oxo-l-proline were increased in these subjects. We were unable to differentiate the patients from each cohort according to their response using both PCA and PLS-DA. However, in SpA patients, five metabolites with a VIP-score >2 from the PLS-DA analysis including PAF-C16, a platelet activating factor, and LysoPC(18:0), a component of cell membranes, appeared relevant. Various metabolic pathways were identified in both studies: glycerophospholipid metabolism; arginine and proline metabolism; metabolism of alanine, aspartate, and glutamate; biosynthesis of phenylalanine, tyrosine, and tryptophan; cysteine and methionine metabolism; biosynthesis of pantothenate and CoA, and biosynthesis of aminoacyl-tRNA.

Conclusion: In patients with RA and SpA, adalimumab induced early changes in the metabolome involving pathways such as essential and non-essential amino acid metabolism and oxidative stress. This finding may guide future investigations to find predictive markers of therapeutic response to TNF inhibitors in chronic inflammatory rheumatic diseases.

Keywords: metabolomics; spondylarthritis; rheumatoid arthritis; adalimumab; response prediction; LC/HRMS; amino acids; oxidative stress.

Introduction

Spondylarthritis (SpA) and rheumatoid arthritis (RA) are the two most common chronic inflammatory rheumatic diseases, affecting 0.32 to 1.4% and 0.5 to 1% of the population, respectively, with regional particularities (1,5). These pathologies induce an important functional impact and an alteration of the quality of life (30–33) as a result from an inflammatory process involving cytokines such as Tumor Necrosis Factor (TNF) alpha. Adalimumab, a recombinant human monoclonal antibody to TNF, is one of the first molecules to be developed and to have proven its efficacy in both diseases (34–37).

However, despite the availability of therapeutic options and a better knowledge of these diseases, an insufficient clinical response persists in approximately one third of patients treated with TNF inhibitors (20–22). Several factors have been mentioned to explain this disparity in response to treatment between patients such as smoking, female sex (38,39), advanced age (20), or the presence of antibodies against treatment (21). Moreover, a poorer response to TNF inhibitors in obese SpA patients has been reported by Micheroli *et al.* (40) and was recently confirmed in patients with RA by Law-Wan J, *et al.* in a meta-analysis (41).

Metabolomics is the study of all metabolites present in living organisms. Metabolites are low molecular weight molecules, involved in multiple processes, as intermediates or final products. The study of pathophysiology through various samples (serum, urine, synovial fluid, stool) from patients suffering from SpA and RA via this field of exploration has allowed the identification of metabolic specific signatures (23,24), essentially in a diagnostic approach given the relatively long time between the onset of symptoms and diagnosis, frequently found in chronic inflammatory rheumatic diseases (42–44).

The contributions of this technique in a process of prediction of response to treatment are still recent although encouraging. Indeed, the analysis of the modification of the metabolome in response to TNF inhibitors has demonstrated the restoration of the balance of certain metabolites in serum in patients with SpA on the one hand and the possibility of differentiating responders from non-responders according to their urinary metabolic profile before treatment in those with RA on the other hand (27,28). Disturbances in lipid metabolism with regulation after TNF inhibitor were identified after analysis of serum, urine, and stool samples from patients with Crohn's disease, suggesting their use as predictive markers of therapeutic response (45).

However, the numbers of these studies are limited with a multiplicity of substrates and a variety of analytical methods that could lead to contradictory results, as found in the literature. So, no definite predictive marker of response to TNF inhibitors has been found so far. Thus, through a post-hoc analysis of serum samples from patients with SpA and RA, our study was to identify possible markers associated with response to adalimumab treatment at 12 and 26 weeks in these two populations, according to the early variation of their metabolome after treatment initiation.

Material and methods

1. Study plan

MetCOMARIS is a post-hoc study of the COMARIS study (Effect of the Combination of Methotrexate and Adalimumab on Reduction of Immunization in Ankylosing Spondylitis; NCT01895764) that was conducted within the HUGO (Hôpitaux Universitaires du Grand Ouest - Western France University Hospitals) network from March 2013 to October 2014. The primary objective was to study the effect of the combination of methotrexate with adalimumab on the immunization of patients with axial SpA. A hundred and ten patients were included.

Patients were randomized in a 1:1 ratio to receive either adalimumab 40 mg subcutaneous injection every two weeks as monotherapy; or in combination with 10 mg weekly subcutaneous injection. This was a prospective, multicentre study that lasted 26 weeks. Results were published in 2020 by Ducourau E, et al. (29).

MetAFORA is a post-hoc study of AFORA (Serum Concentration of Adalimumab as a Predictive Factor of clinical Outcomes in Rheumatoid Arthritis; NCT01382160) which was also conducted within the HUGO network and whose main objective was to estimate the pharmacodynamic parameters of the concentration-effect relationship of adalimumab in RA. AFORA was conducted from January 2011 to January 2013 with a 26-week follow-up of patients. The results of this study are still under analysis.

In the present study, we performed metabolomic analyses on samples from patients in each cohort according to their clinical response to adalimumab treatment, to assess early metabolome changes potentially involved in the response to treatment.

2. Detailed study methodology

a. Patients

The following inclusion criteria were similar in both cohorts:

- Male or female of age greater than or equal to 18 years.
- With ASAS diagnostic criteria of axial SpA or ACR (1987) diagnostic criteria of RA (**Annexes I**).
- Justifying treatment with adalimumab in accordance with the marketing authorization.

More specifically to the COMARIS cohort:

- Absence of methotrexate administration in the last 3 months and the absence of previous treatment with a TNF inhibitor.

More specifically to the AFORA cohort:

- Stability of background methotrexate and/or corticosteroid therapy 4 weeks prior to inclusion and at least during the first 12 weeks of the study.

b. Clinical data

Levels of spinal pain, peripheral pain and swelling, and duration of morning stiffness were collected using a visual analogue scale at each visit (W4, W12 and W26) in the COMARIS cohort. A joint index, synovitis and patients' assessment of disease activity using a visual analogue scale were reported at each visit (W4, W12 and W26) in the AFORA cohort.

All these clinical elements associated with the biological data were used to calculate the activity score specific to each cohort: ASDAS in the COMARIS cohort and DAS28 in the AFORA cohort. According to the ASDAS score obtained, SpA was considered as 'inactive', 'moderately active', 'active', or 'very active'. Similarly, according to the DAS28 score obtained, RA was in 'remission', 'low', 'moderate', or 'high' activity (**Annexes II**).

c. Blood samples and biological data

Blood samples were taken before adalimumab injections and then at each visit at W4, W12 and W26. The samples were centrifuged, aliquoted, labelled and stored in each investigating centre before being sent centrally in dry ice to the Tours centre where all samples were stored at -80°C. The erythrocyte sedimentation rate (ESR) and C-reactive protein (CRP) were measured in the biochemistry laboratories of the Centre Hospitalier Régional Universitaire de Tours under the responsibility of Eric Piver.

d. Evaluation of the therapeutic response

Therapeutic response was defined according to the change in each activity score between inclusion and the 12- and 26-week clinical assessment.

In the COMARIS cohort, based on the degree of change in ASDAS, therapeutic response was defined as: 'absent'; 'significant' or 'major'. In the AFORA cohort, according to the variation of DAS28, therapeutic response was defined as: 'absent'; 'moderate' or 'good' (**Annexes III**).

In our study, we chose to group patients into "responders" and "non-responders" according to the following modalities:

- COMARIS cohort: those with a 'significant' or 'major' response were considered "responders" and those with 'absent' were considered "non-responders".
- AFORA cohort: those with a 'moderate' or 'good' response are considered "responders" and those with 'absent' were considered "non-responders".

e. Metabolomic data

One-millilitre serum samples were analysed. They were first thawed for 1 hour 30 minutes and then 50 µL of each serum was collected. The metabolites were extracted with 400µL of methanol added to 50µL of serum. The samples were vortexed for 5 seconds and then incubated at -20°C for 30 minutes to deproteinize the sample. After centrifugation for 25 min at 5000 rpm at 4°C, the supernatant (350 µL) was harvested and transferred to a 96-well plate. After evaporation of the plate under nitrogen, the dry residue was resuspended in 100 µL of methanol/water (75/25) for C18 analysis, in 100 µL of acetonitrile/water (75/25) for hydrophilic interaction liquid chromatography (HILIC) analysis. After vortexing again for 20 seconds, the supernatant was transferred to a 500 µL Eppendorf tube. The tube was centrifuged for 15 minutes at 5000 rpm at 4°C. Five µL of sample was injected for analysis by positive and negative ionization mass spectrometry. The analytical technique was liquid chromatography coupled to high-resolution mass spectrometry (LC/HRMS). The analyses were performed on an Ultimate WPS-3000 UPLC system (Dionex, Germany) coupled to a Q-Exactive mass spectrometer (Thermo Fisher Scientific, Bremen, Germany).

Liquid chromatography was performed using two different modes:

- HILIC to analyse polar molecules with HILIC Cortecs columns (1.6µm 150x2.10m) maintained at 40°C. Two mobile phases were used, and chromatographic gradients were performed at a flow rate of 0.3mL performed at a flow rate of 0.3mL / min.
- C18 to analyse apolar molecule. C18xb columns (1.7µm 150 x 2.1mm) were maintained at 55°C. Two mobile phases were used, and chromatographic gradients were performed at a flow rate of 0.3mL/min.

Ionization of the molecules was performed using the positive (ESI+) and negative (ESI-) electrospray method. During acquisition, ions were scanned from 58 to 870 m/z at a resolution of 70,000 (m/z = 200). The analyses were processed in a so-called "targeted" manner. A library of 530 molecules has already been characterized with the chromatographic conditions used for this study (C18 and HILIC combined). Signal values were calculated with the Thermo Xcalibur processing configuration software (Thermo Fisher Scientific) by integrating the chromatography peaks corresponding to the selected metabolites. Once the Excel data table with the metabolites was obtained, different data pre-treatments were performed. Analyses were performed by Antoine Lefevre under the supervision of Patrick Emond.

In case the metabolite was not detectable, the lowest measured value within the cohort was used as a surrogate to calculate the variation between the inclusion visit and W4.

3. Statistical analysis

Clinical and biological data from AFORA and COMARIS cohorts at inclusion were analysed using BiostaTGV. Quantitative values were analysed with a student's t test and qualitative variables with a Fisher's test. The significance level was set at 0.05.

Metabolomic profiles of patients from both cohorts were independently analyzed with the online software METABOANALYST 5.0., at baseline and then between W0 and W4 depending on the therapeutic response at 12 weeks (primary objective) and 26 weeks (secondary objective).

First, we performed a univariate analysis, followed by an unsupervised principal component analysis (PCA). Then we performed a partial least squares discriminant analysis (PLS-DA), a supervised method of multiple linear regression to separate the data of each cohort into two groups to find the maximum covariance at baseline and between W0 and W4 according to the therapeutic response at 12 and 26 weeks.

Permutation tests were performed to verify the reliability of our models and then we identified variables of interest, defined as those with a VIP (Variable Influence on Projection) score greater than or equal to 2, from the PLS-DA analysis in each cohort.

From these variables of interest, we proceeded to the analysis of the metabolic pathways involved at the inclusion, then in the early variation of the metabolome according to the therapeutic response at 12 and 26 weeks in each cohort. Those found in the early variation of the metabolome according to the therapeutic response at 12 weeks were then compared between the two cohorts.

Those analysis were performed by Elom Tay under the supervision of Hélène Blasco.

Results

1. Population characteristics

In the COMARIS cohort, at 12 weeks, we excluded 30 patients with no metabolomic data available over the entire study period, 1 patient for whom the ASDAS was missing, 2 patients for whom the metabolomic data at inclusion were missing and 3 patients for whom the clinical response data was missing. At 26 weeks, we excluded 1 patient for whom the ASDAS score at inclusion was missing, 7 patients for whom the clinical response was missing and 1 patient for whom the metabolomic data at inclusion was missing. We therefore studied the sera of 74 patients in the primary analysis and 71 patients in the secondary analysis.

In the AFORA cohort, 69 patients were included. The results of 2 patients could not be studied, 2 patients left the study at 4 weeks due to lack of clinical response, one patient was wrongly included, and we had a duplicate of data for one patient at 4 weeks. In addition, at 26 weeks, clinical response for 7 patients were lacking. We therefore studied the sera of 63 patients in the primary analysis and 56 patients in the secondary analysis.

All these data are shown in **Figure 1**.

In the COMARIS cohort, 69.76% of responders were male ($p = 0.002$). There was no significant difference in age or body mass index between the two groups. CRP was significantly increased at baseline in responders ($p = 0.0002$) and both groups had active disease ($p = 0.002$).

In the predominantly female AFORA cohort, all men appeared to be responders ($p = 0.053$). There was no significant difference by age, CRP or DAS28. Responders were borderline overweight ($p = 0.049$).

Finally, there was no difference in the association with methotrexate between responders and non-responders in the two cohorts. These results are shown in **Table 1**.

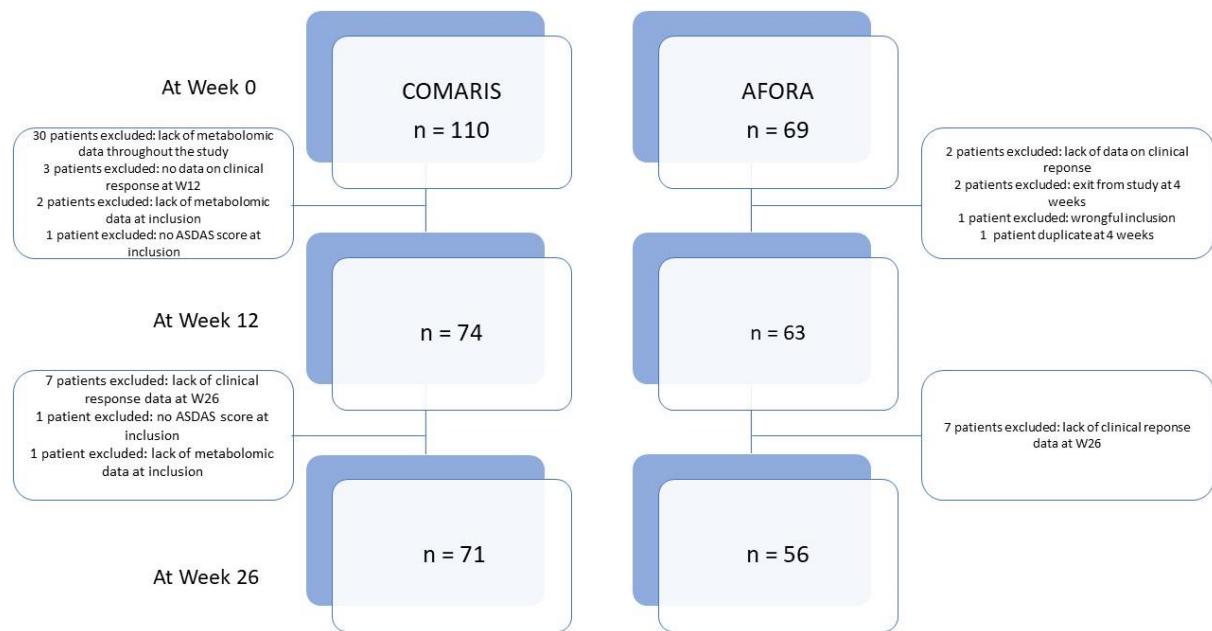


Figure 1. Flow chart of the COMARIS and AFORA cohorts.

Table 1. Baseline characteristics by adalimumab response at 12 Weeks (COMARIS n = 74; AFORA n = 63)

Cohort	COMARIS			AFORA		
Clinical status at 12 weeks	Responders (n= 43)	Non-responders (n= 31)	p-value	Responders (n= 52)	Non-responders (n= 11)	p-value
Male (n, %)	30 (69,76%)	10 (32,25%)	0,002	15 (28%)	0	0,053
Age, mean (years)	41,25	32,25	0,197	54,88	53,36	0,746
BMI, mean (kg/m ²)	25,81	25,31	0,623	25,65	23,03	0,049
CRP (mg/l)	13,58	3,22	0,0002	10,87	16,93	0,348
ASDAS-CRP/DAS28	3,42	2,83	0,002	4,8	5,19	0,398
MTX (n, %)	19 (44,18%)	14 (45,16%)	1	35 (67,30%)	7 (63,63%)	1
Corticosteroids	-	-		25 (48,07%)	5 (45,45%)	1

2. Metabolomic analyses

2.1. COMARIS

- Main analysis: based on therapeutic response at 12 weeks
 - Metabolome analysis at W0 for response prediction

We were not able to separate the metabolomic profiles of patients according to their therapeutic response by either PCA or PLS-DA. There were 2 outliers in PCA and 3 in PLS-DA (**Annexes IV**). The resulting model can only be considered descriptive ($p = 0.46$ by permutation test). These analyses revealed higher concentrations of thymidine and theobromine in responders and lower concentrations of L-alanine, L-pipecolic_acid, N-acetyl-l-alanine, sarcosine, methyl_indole-3-acetate, and c4-carnitine (VIP-score ≥ 2) (**Annexes, Figure 1**).

The following metabolic pathways could be identified: linoleic acid metabolism; glycine, serine, and threonine metabolism; pyrimidine metabolism; and arginine and proline metabolism (**Annexes, Figure 2**).

- Analysis of the metabolome change between W0 and W4

We were not able to separate the metabolomic profiles of patients according to their therapeutic response at 12 weeks neither in PCA nor in PLS-DA, **Figure 2**. The permutation test showed a p value of 0.65.

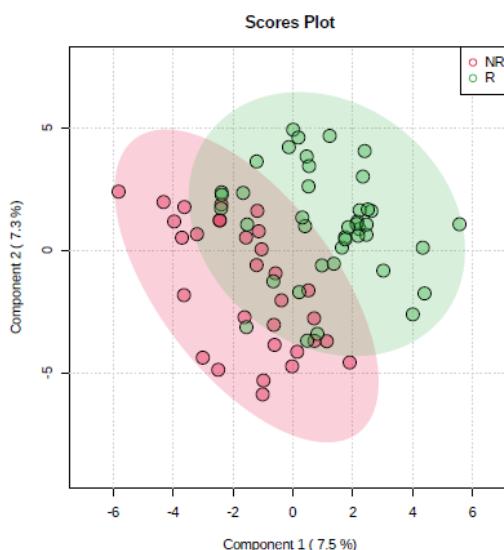


Figure 2. COMARIS. PLS-DA separation of different patient groups according to metabolome change between W0 and W4 based on clinical response at 12 weeks. R = responders; NR = non-responders.

We highlighted 15 metabolites of interest including 5 with a VIP-score ≥ 2 that are: 5,6-dihydrouracil, l-cystine, and retinoate increased in responders whereas LPC(18:0) (LyoPC 18:0) and PAF-C16 (Platelet Activating Factor - C16) were decreased in this subgroup, **Figure 3**.

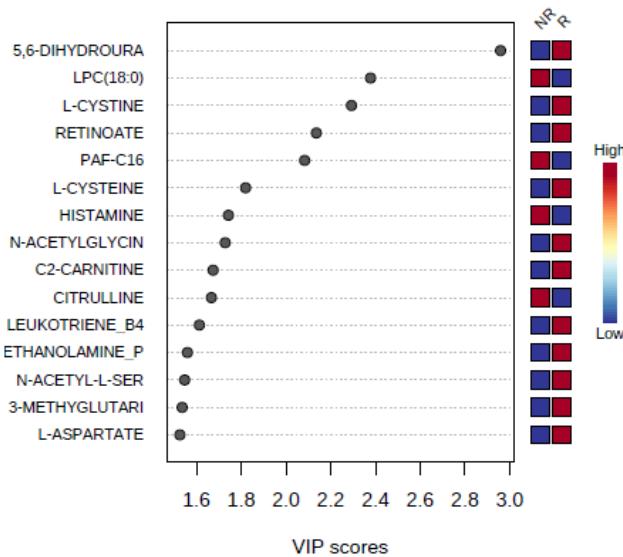


Figure 3. COMARIS. Score-plot of metabolites of interest of different patient groups according to metabolome change between W0 and W4 as a function of clinical response at 12 weeks. R = responders; NR = non-responders.

We were able to identify the following metabolic pathways: arginine biosynthesis; retinol metabolism; alanine, aspartate, and glutamate metabolism; histidine metabolism; cysteine and methionine metabolism; beta-alanine metabolism; and pantothenate and CoA biosynthesis, **Figure 4**.

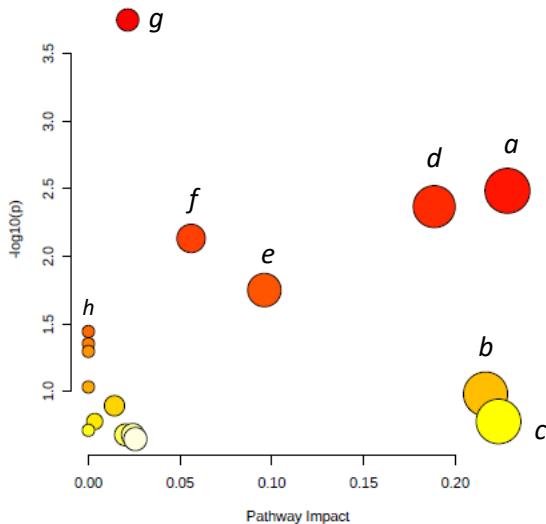


Figure 4. COMARIS. Metabolic pathways involved in the metabolome change between W0 and W4 according to the therapeutic response at 12 weeks. a: arginine biosynthesis; b: retinol metabolism; c: alanine, aspartate, and glutamate metabolism; d: histidine metabolism; e: cysteine and methionine metabolism; f: beta-alanine metabolism; g: pantothenate and CoA biosynthesis; h: aminoacyl-tRNA biosynthesis.

- b. Secondary analysis: based on therapeutic response at 26 weeks
 - Metabolome analysis at W0 for response prediction

We did not observe a separation in PCA, nor in PLS-DA between the two groups ($p = 0.53$ by permutation test). There were 3 outliers in both analyses (**Annexes IV**). Metabolites such as allantoin, theobromine, and cholestryl acid appeared increased in responders, whereas trans-cinnamate was lowered in these subjects (VIP-score ≥ 2) (**Annexes, Figure 3**).

The metabolic pathways that could be identified were glycine, serine, and threonine metabolism; phenylalanine, tyrosine, and tryptophan biosynthesis; tyrosine metabolism;

arginine and proline metabolism; taurine and hypotaurine metabolism; beta-alanine metabolism; and histidine metabolism. (**Annexes, Figure 4**).

- Analysis of the metabolome change between W0 and W4

Analyses in PCA and PLS-DA did not separate metabolomic profiles by response status ($p = 0.73$ by permutation test). There was 1 outlier in PLS-DA (**Annexes IV**). Among the metabolites of interest from PLS-DA analysis, we found PAF-C16 and LPC (18:0), both lowered in responders. They are shown in (**Figure 5**) in the (**Annexes**).

Metabolic pathway analysis identified the following pathways: aminoacyl-tRNA biosynthesis; valine, leucine, and isoleucine metabolism; glycine, serine, and threonine metabolism; glutathione metabolism; glyoxylate and dicarboxylate metabolism; arginine and proline metabolism; d-glutamine and glutamate metabolism; and alanine, aspartate, and glutamate metabolism (**Annexes, Figure 6**).

c. Common metabolic pathways

Metabolites such as PAF-C16 and LPC (18:0) appear to be the only relevant components of the metabolome variation between W0 and W4 between responders and non-responders. Metabolic pathway analysis revealed a few common pathways at both time points represented in **Figure 5**.

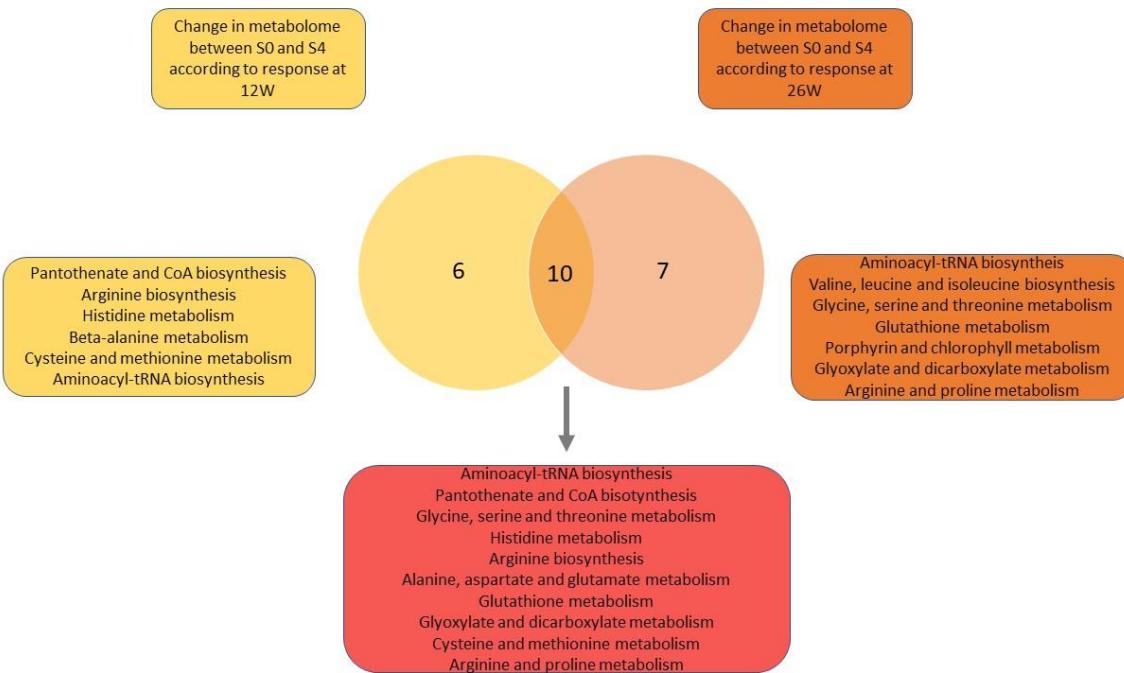


Figure 5. COMARIS. Venn diagram representing common metabolic pathways in metabolome analyses between W0 and W4 according to therapeutic responses at 12 and 26 weeks

2.2.AFORA

- a. Main analysis: based on therapeutic response at 12 weeks
 - Metabolome analysis at W0 for response prediction

The analysis in PCA did not allow us to separate the metabolomic profiles of patients according to their therapeutic response. This separation was possible in PLS-DA but with a $p = 0.70$ in the permutation test (**Annexes, Figure 7**). There were 4 outliers in PCA and 3 in PLS-DA (**Annexes IV**).

The metabolites of interest are shown in (**Figure 8**) in the (**Annexes**). Those with a VIP-score ≥ 2 are paraxanthin, theobromine, catechol, pantothenic acid, n-acetylglycine, o-acetyl-l-carnitine, rac-glycerol_1 and lauroylcarnitine.

Among the metabolic pathways possibly involved at W0, we identified: caffeine metabolism; ketone body synthesis and degradation; and butanoate metabolism (**Annexes, Figure 9**).

- Analysis of the metabolome change between W0 and W4

The metabolomic profiles of the patients could not be separated by either PCA or PLS-DA analysis ($p = 0.23$ by permutation test) as shown in **Figure 6**. There were 4 outliers in PLS-DA (**Annexes IV**).

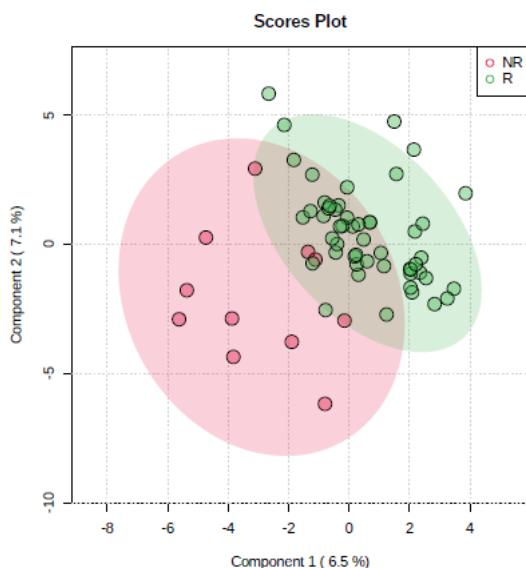


Figure 6. AFORA. PLS-DA separation of different patient groups according to metabolome change between W0 and W4 based on clinical response at 12 weeks. R = responders; NR = non-responders

15 metabolites of interest could be identified in PLS-DA, including 7 with a VIP-score ≥ 2 : carnosine; l-ornithine; l-valine; l-glutamic_acid; n-acetyl-serine; l-serine; l-threonine, all of which were lowered in responders, **Figure 7**.

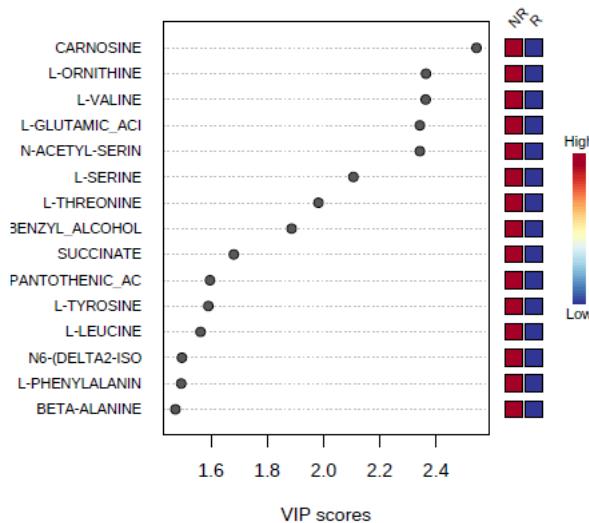


Figure 7. AFORA. Score-plot of metabolites of interest of different patient groups according to metabolome change between W0 and W4 according to clinical response at 12 weeks. R = responders; NR = non-responders.

Part of the highlighted metabolic pathways are shown in **Figure 8**, to which are added the pathways of butanoate metabolism; histidine metabolism; propanoate metabolism; glutathione metabolism and glyoxylate and dicarboxylate metabolism.

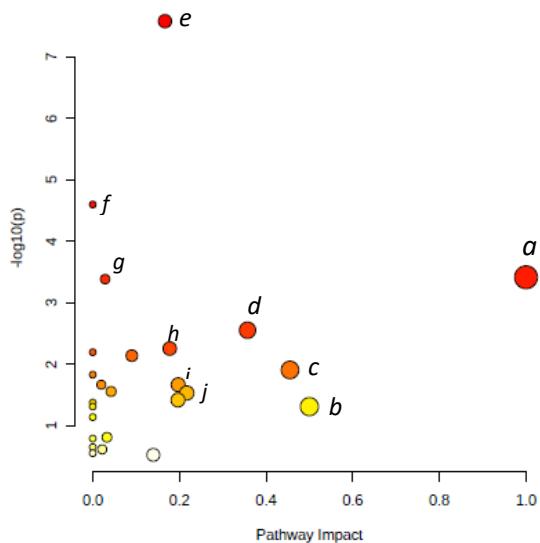


Figure 8. AFORA. Metabolic pathways involved in the metabolome change between W0 and W4 according to therapeutic response at 12 weeks. a: phenylalanine, tyrosine, and tryptophan biosynthesis; b: D-glutamine and D-glutamate metabolism; c: beta-alanine metabolism; d: phenylalanine metabolism; e: Aminoacyl-t-RNA biosynthesis; f: valine, leucine, and isoleucine biosynthesis; g: pantothenate and CoA biosynthesis; h: arginine biosynthesis; i: alanine, aspartate, and glutamate metabolism; j: glycine, serine, and threonine metabolism.

- b. Secondary analysis: based on therapeutic response at 26 weeks
 - Metabolome analysis at W0 for response prediction

The two groups of patients could be correctly separated by the PLS-DA analysis although it included 2 outliers (**Annexes IV**). However, these results can only be interpreted as descriptive due to a p value = 1 in the permutation test. Of the 15 metabolites identified, 6 had a VIP-score ≥ 2 and were increased in responders: l-lysine; 5-aminopentanoate; glyceraldehyde; l-threonine; guanine; and l-phenylalanine (**Annexes, Figure 10**).

The metabolic pathways involved at this stage are shown in (Figure 11) in the (Annexes), including aminoacyl-tRNA biosynthesis; lysine degradation; glycine, serine, and threonine metabolism; and phenylalanine, tyrosine, and tryptophan biosynthesis.

- Analysis of the metabolome change between W0 and W4

There was a slight overlap between the two groups in PLS-DA analysis. The metabolites of interest are shown in (Figure 12) in the (Annexes). Gluconic acid, n-acetylglycine, and glycine were all increased in responders with a VIP-score ≥ 2 .

The metabolic pathways that could be demonstrated were the pentose phosphate pathway; glutathione metabolism; glyoxylate and dicarboxylate metabolism; glycine, serine, and threonine metabolism; and arginine biosynthesis. They are shown in (Figure 13) in the (Annexes).

c. Common metabolic pathways

We were not able to identify metabolites whose variation over time appeared to be related to therapeutic response, but several metabolic pathways could be identified, shown in **Figure 9** below.

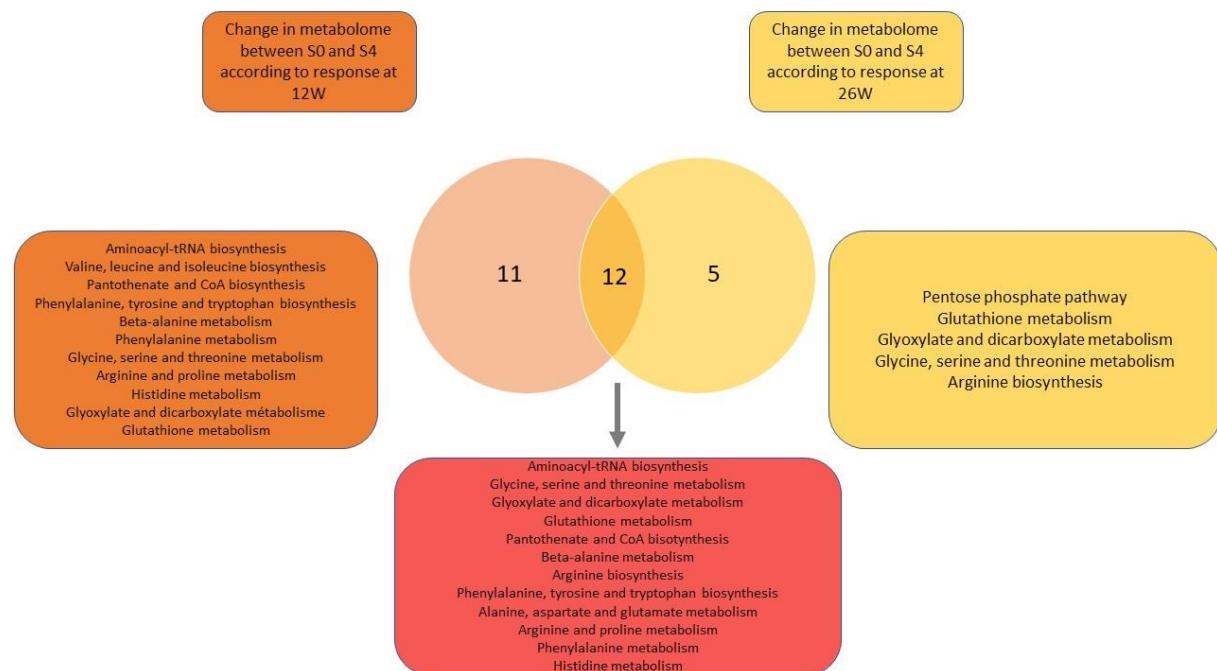


Figure 9. AFORA. Venn diagram representing common metabolic pathways for metabolome analyses between W0 and W4 according to therapeutic responses at 12 and 26 weeks.

2.3.Comparison of the two cohorts

The pathways that appear to be involved in early metabolome variation in response to adalimumab treatment in both cohorts are shown in **Figure 10**.

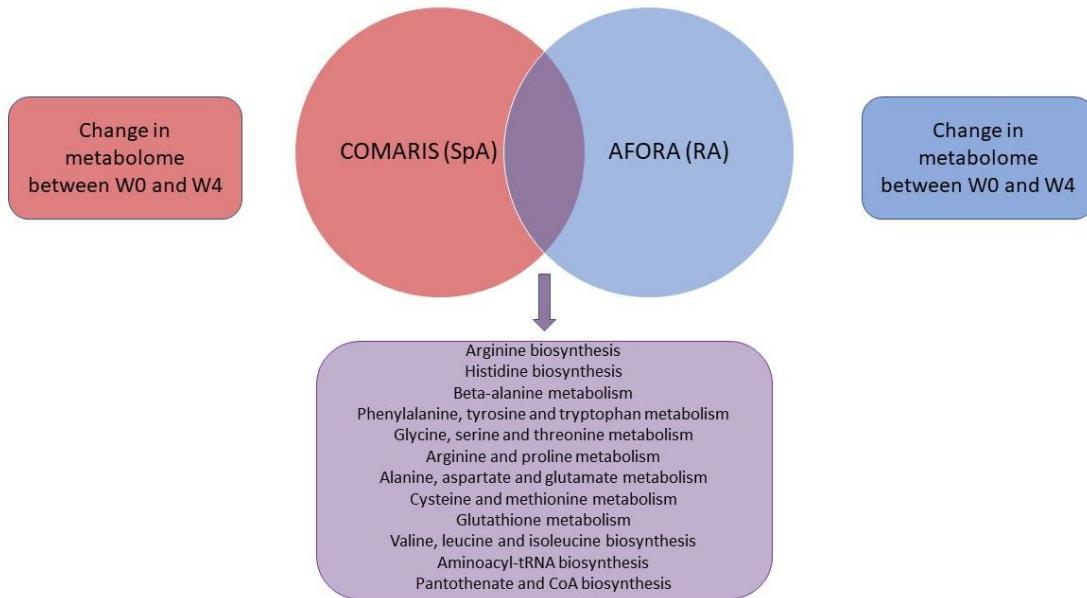


Figure 10. Metabolic pathways common to both cohorts based on metabolome change between W0 and W4.

Discussion

In our study, we tried to identify metabolites whose early variation could differentiate responders from non-responders to adalimumab treatment in patients with SpA and RA by analysing their metabolomic profiles by LC/HRMS. Therapeutic responses were collected at 12 and 26 weeks (respectively primary and secondary endpoints) based on the different clinical activity scores of these conditions in accordance with the recommendations of scientific societies. The characteristics of our model allow us to consider it only as descriptive without the ability to differentiate patients according to their clinical response. However, we were able to highlight relevant metabolites and metabolic pathways already well described in the literature of these two diseases in relation with clinical response, such as the metabolism of glycine, serine, and threonine; arginine and proline metabolism; the metabolism of alanine, aspartate, and glutamate; biosynthesis of phenylalanine, tyrosine, and tryptophan; biosynthesis of valine, leucine, and isoleucine; cysteine and methionine metabolism; glutathione metabolism and glycerophospholipid metabolism.

Metabolome analysis at treatment initiation according to therapeutic response at 12 weeks in the COMARIS cohort showed high levels of thymidine and theobromine in responders, while the following metabolites: l-alanine, l-pipecolic_acid, n-acetyl-l-alanine, sarcosine, methyl_indole-3-acetate, and carnitine were lowered. Low serum carnitine concentrations were indeed found in subjects with SpA (24,26) while alanine appeared to be increased in these patients (26,46). We did not find any data concerning thymidine and theobromine in the literature, nor on the possible effect of TNF inhibitors on the regulation of the concentrations of all these metabolites.

When studying the variation of the metabolome between W0 and W4 in the COMARIS cohort, we noticed a decrease in PAF-C16 and LysoPC (18:0) concentrations in relation with the therapeutic response at 12 and 26 weeks. PAF-C16 (also known as PAF) is a ubiquitous molecule belonging to the phospholipid family that acts as a potent activator and mediator in inflammatory processes and cardiovascular diseases. Its involvement in autoimmune diseases, linked or not to its receptor (PAF receptor) was reported by Edwards LJ *et al.* in a review where its concentration was increased in the intestinal mucosa and stool of patients with inflammatory bowel diseases compared to healthy patients (47).

In its storage form, lipo-PAF, it appeared to be higher in patients with inflammatory rheumatic diseases and in those with osteoarthritis or chondrocalcinoses compared to healthy subjects with the particularity of being also elevated in the synovial fluid of patients with RA (47) (48). An interdependence between TNF and PAF has been demonstrated in numerous studies, the former acting at the level of endothelial cells to induce PAF synthesis (49,50); the second acting on the peripheral monocytes resulting in a production of TNF (51). The consequence was a synergistic inflammatory action at the synovial level (52,53). Thus, PAF appears to be lowered in responders to TNF inhibitors.

LysoPC(18:0) is a lysophospholipid, belonging to the family of lysophosphatidylcholines, molecules involved in fatty acid metabolism. It is present in small quantities in many tissues. An elevated concentration of LysoPC (18:0) was found in patients with SpA compared to healthy subjects without modification after treatment with TNF inhibitor. High concentrations of this metabolite could also be found in the stool of patients with rheumatoid arthritis compared to healthy subjects, supporting the disruption of glycerophospholipid metabolism in these two conditions (27,54).

Interestingly, retinoate was one of the metabolites of interest in the study of metabolome variation between W0 and W4 in relation to therapeutic response at 12 weeks in the COMARIS cohort. It was increased in responders and is known as all-trans-retinoic acid (ATRA), an important regulator of gene expression during growth. Retinoids are known to be involved in bone formation and structure, and in patients with SpA, in whom retinol concentration was disturbed (55). In addition, the *in vitro* study of the action of ATRA, an active metabolite of vitamin A, on T lymphocytes made it possible to identify its regulatory action on the pathway of Th17 and TNF-alpha synthesis, which were lowered in the cultures studied after treatment (56). Finally, Wu J. et al. were able to demonstrate the effect of adalimumab in restoring the concentration of certain proteins such as Retinol Binding Protein 4 (RBP4), whose increase could be used as a predictive marker of a good response to treatment in patients with ankylosing spondylitis (57).

In our RA cohort, metabolome analysis at W0 according to therapeutic response at 12 weeks found high levels of paraxanthin, theobromine, glyceraldehyde, catechol, n-acetylglycine, pantothenic acid, o-acetyl-l-carnitine, and rac-glycerol_1 in responders. Pantothenic acid was described in the 1960s as a potential marker for RA but this has not been confirmed by further studies. Glyceraldehyde is known to be increased in patients with RA (24,25). High levels of xanthine have been found in the urine of patients treated with TNF inhibitors (58); n-acetylglycine was found in increased concentration in patients with RA compared to healthy subjects in numerous studies (58) and was increased in etanercept responders in the study by Priori et al. (59). Glycerol is also known to be increased in patients with RA (60–62) with a decrease in glycerol 3 phosphate after 12 weeks of treatment with a TNF inhibitor (63). Finally, carnitine appears to be lowered in patients with RA (24).

Analysis of the metabolome variation between W0 and W4 according to the therapeutic response at 12 weeks did not identify relevant metabolites. Only n-acetylglycine appeared to be increased in responders according to therapeutic response at 26 weeks, consistent with the data from the study by Priori et al. (59)

Disturbances in amino acid metabolism have been well described in SpA and RA. A strong association between the combination of concentrations of phenylalanine, tryptophan, threonine, aspartic acid, histidine and the number of painful, swollen joints as well as DAS28 in RA has been reported by Smolenska et al. (64). Serine, proline and alanine, non-essential amino acids, have been described as associated with SpA activity (26). More specifically, proline, an

essential element in the synthesis of many human proteins, has been found to be increased in both diseases. Dysfunctions of its metabolism can induce abnormalities in the citric acid cycle, disrupting cellular metabolism and can cause an inflammatory state damaging cartilage and bone (25,26). Its synthesis involves arginine, described as increased in subjects with SpA and rather decreased in those with RA (25,26).

Arginine is needed for the synthesis of nitric oxide, involved in the functioning of endothelial cells with consequences on atherosclerosis and the development of cardiovascular pathologies when this pathway is disrupted. This phenomenon has already been studied in inflammatory rheumatic diseases with encouraging results (65–67). Changes in the metabolic pathways of glycine, serine, and threonine as well as arginine and proline have been reported by Kamleh et al. after treatment with etanercept in patients with psoriasis where it was observed a return to the equilibrium state of these pathways (68).

Ou et al. described glutamate as being significantly increased in serum in patients with SpA with a strong correlation with disease activity. It is an amino acid derived from glutaminolysis involved in the production of energy for cells including T lymphocytes with a favor of the development of the Th17 pathway. The regulation of its metabolism after treatment with TNF inhibitor was observed in their study (27). In addition, data exist on the key role of glutaminolysis in the cell growth of fibroblast-like synoviocytes in RA (69). The decrease in glutamate after treatment was also found by Gupta et al. in patients with SpA (46).

High concentrations of valine, leucine and isoleucine, essential amino acids, have been found in patients with SpA, with a recovery of balance of their metabolism after treatment (27,46). Disturbances in the concentration of phenylalanine were found in both diseases and were influenced by TNF inhibitors (25,27). Low concentrations of tryptophan, another essential amino acid, have been reported in both diseases with a correlation between disease activity in SpA and the breakdown of the bond between tryptophan and its binding protein (25,70). In addition, recent studies have demonstrated some disruptions in tryptophan metabolism in connection with dysregulation of the intestinal microbiota in both SpA and RA patients (54,71).

Cysteine and its precursor, methionine are involved in oxidative stress metabolism with the participation of glutathione. Oxygen and nitrogen reactive species are known to participate in acute or chronic inflammatory processes by interacting with proteins, lipids and DNA thus being able to generate autoantibodies such as citrullinated cyclic peptide antibodies in RA. An increase in these reactive species at the synovial level in RA has also been reported, resulting in local hypoxia and joint destruction (23,72–74). In patients with SpA, a hostile serum microenvironment containing inflammatory markers and advanced oxidative protein products with deleterious action (senescence and apoptosis) on mesenchymal stem cells known to have immunoregulation properties has been reported (75). These advanced oxidative protein products are also known to be linked to disease activity (76). TNF alpha is known to be involved in oxidative stress by inducing the inappropriate production of nitric oxide, a highly reactive species (77). The anti-oxidant effect of TNF inhibitors could be demonstrated by Yamamoto K et al. in patients with Crohn's disease treated with infliximab (another TNF inhibitor) and adalimumab without inducing the formation of antioxidant molecules (78).

The main strength of our study is its longitudinal methodology and the novelty of such metabolomic analyses coupled to the clinic for predictive purposes on relatively large samples. We were able to study the metabolomic profile of a total of 137 patients with the two most common inflammatory rheumatic diseases. In addition, we were able to maintain a certain homogeneity of treatment by using the same TNF inhibitor, namely adalimumab.

However, it also has some limitations. Owing to its post-hoc design, blood samples were not intended for metabolomic analyses. As a result, transport and storage conditions have not been standardized. In addition, we used serum as a substrate and not plasma in which the blood clots, a mechanism involving the metabolism of glycerophospholipids (79). We did not control for other factors that may alter patients' metabolism such as diet or cardiovascular comorbidities (dyslipidemia, high blood pressure, obesity/sarcopenia) which is common in patients with inflammatory rheumatic diseases (80,81). Finally, we did not have a unisex population as specific metabolomic profiles by sex were described in the literature both in general population and inflammatory rheumatic diseases (82–84).

Conclusion

Our study aimed to identify metabolites associated with early response to adalimumab treatment in patients with SpA and RA, the two most common inflammatory rheumatic diseases. We were able to identify relevant elements aligned with current data in the literature regarding the metabolism of amino acids (essential and non-essential) and oxidative stress that could serve as a basis for future investigations in the search for predictive markers of response to treatment.

References

1. Sieper J, Poddubny D. Axial spondyloarthritis. *The Lancet*. 1 juill 2017;390(10089):73-84.
2. Sharip A, Kunz J. Understanding the Pathogenesis of Spondyloarthritis. *Biomolecules*. 20 oct 2020;10(10):1461.
3. Maksymowich WP. Les biomarqueurs dans la spondylarthrite : de la physiopathologie à l'évaluation de la maladie. *Rev Rhum*. 1 mars 2012;79(2):101-3.
4. Otón T, Carmona L. The epidemiology of established rheumatoid arthritis. *Best Pract Res Clin Rheumatol*. 1 oct 2019;33(5):101477.
5. Roux CH. Impact of musculoskeletal disorders on quality of life: an inception cohort study. *Ann Rheum Dis*. 1 avr 2005;64(4):606-11.
6. Silman AJ, Newman J, MacGregor AJ. Cigarette smoking increases the risk of rheumatoid arthritis. Results from a nationwide study of disease-discordant twins. *Arthritis Rheum*. mai 1996;39(5):732-5.
7. Klareskog L, Malmström V, Lundberg K, Padyukov L, Alfredsson L. Smoking, citrullination and genetic variability in the immunopathogenesis of rheumatoid arthritis. *Semin Immunol*. 1 avr 2011;23(2):92-8.
8. Manasson J, Blank RB, Scher JU. The microbiome in rheumatology: Where are we and where should we go? *Ann Rheum Dis*. juin 2020;79(6):727-33.
9. Smolen JS, Aletaha D, McInnes IB. Rheumatoid arthritis. *The Lancet*. oct 2016;388(10055):2023-38.
10. Lukas C, Landewé R, Sieper J, Dougados M, Davis J, Braun J, et al. Development of an ASAS-endorsed disease activity score (ASDAS) in patients with ankylosing spondylitis. *Ann Rheum Dis*. janv 2009;68(1):18-24.
11. van der Heijde D, Lie E, Kvien TK, Sieper J, Van den Bosch F, Listing J, et al. ASDAS, a highly discriminatory ASAS-endorsed disease activity score in patients with ankylosing spondylitis. *Ann Rheum Dis*. déc 2009;68(12):1811-8.
12. Prevoo ML, van 't Hof MA, Kuper HH, van Leeuwen MA, van de Putte LB, van Riel PL. Modified disease activity scores that include twenty-eight-joint counts. Development and validation in a prospective longitudinal study of patients with rheumatoid arthritis. *Arthritis Rheum*. janv 1995;38(1):44-8.
13. Machado P, Landewé R, Lie E, Kvien TK, Braun J, Baker D, et al. Ankylosing Spondylitis Disease Activity Score (ASDAS): defining cut-off values for disease activity states and improvement scores. *Ann Rheum Dis*. janv 2011;70(1):47-53.
14. van Gestel AM, Haagsma CJ, van Riel PL. Validation of rheumatoid arthritis improvement criteria that include simplified joint counts. *Arthritis Rheum*. oct 1998;41(10):1845-50.
15. Fransen J, van Riel PL. DAS remission cut points. *Clin Exp Rheumatol*. déc 2006;24(6 Suppl 43):S-29-32.
16. Smolen JS, van der Heijde D, Machold KP, Aletaha D, Landewé R. Proposal for a new nomenclature of disease-modifying antirheumatic drugs. *Ann Rheum Dis*. janv 2014;73(1):3-5.
17. Sánchez-Piedra C, Sueiro-Delgado D, García-González J, Ros-Vilamajo I, Prior-Español A, Moreno-Ramos MJ, et al. Changes in the use patterns of bDMARDs in patients with rheumatic diseases over the past 13 years. *Sci Rep*. 23 juill 2021;11:15051.

18. Smolen JS, Landewé RBM, Bijlsma JWJ, Burmester GR, Dougados M, Kerschbaumer A, et al. EULAR recommendations for the management of rheumatoid arthritis with synthetic and biological disease-modifying antirheumatic drugs: 2019 update. *Ann Rheum Dis.* juin 2020;79(6):685-99.
19. Ward MM, Deodhar A, Gensler LS, Dubreuil M, Yu D, Khan MA, et al. 2019 Update of the American College of Rheumatology/Spondylitis Association of America/Spondyloarthritis Research and Treatment Network Recommendations for the Treatment of Ankylosing Spondylitis and Nonradiographic Axial Spondyloarthritis. *Arthritis Rheumatol Hoboken NJ.* oct 2019;71(10):1599-613.
20. Hetland ML, Christensen IJ, Tarp U, Dreyer L, Hansen A, Hansen IT, et al. Direct comparison of treatment responses, remission rates, and drug adherence in patients with rheumatoid arthritis treated with adalimumab, etanercept, or infliximab: Results from eight years of surveillance of clinical practice in the nationwide Danish DANBIO registry. *Arthritis Rheum.* janv 2010;62(1):22-32.
21. Juanola X, Ramos MJM, Belzunegui JM, Fernández-Carballedo C, Gratacós J. Treatment Failure in Axial Spondyloarthritis: Insights for a Standardized Definition. *Adv Ther.* 2022;39(4):1490-501.
22. Moral E, Plasencia C, Navarro-Compán V, Pascual Salcedo D, Jurado T, Tornero C, et al. AB0657 Discontinuation of Anti-TNF Therapy in Patients with Axial Spondyloarthritis in Clinical Practice: Prevalence and Causes. *Ann Rheum Dis.* 1 juin 2016;75(Suppl 2):1129.
23. Jutley GS, Young SP. Metabolomics to identify biomarkers and as a predictive tool in inflammatory diseases. *Best Pract Res Clin Rheumatol.* 1 déc 2015;29(6):770-82.
24. Jiang M, Chen T, Feng H, Zhang Y, Li L, Zhao A, et al. Serum metabolic signatures of four types of human arthritis. *J Proteome Res.* 2 août 2013;12(8):3769-79.
25. Li J, Che N, Xu L, Zhang Q, Wang Q, Tan W, et al. LC-MS-based serum metabolomics reveals a distinctive signature in patients with rheumatoid arthritis. *Clin Rheumatol.* juin 2018;37(6):1493-502.
26. Zhou Y, Zhang X, Chen R, Han S, Liu Y, Liu X, et al. Serum amino acid metabolic profiles of ankylosing spondylitis by targeted metabolomics analysis. *Clin Rheumatol.* août 2020;39(8):2325-36.
27. Ou J, Xiao M, Huang Y, Tu L, Chen Z, Cao S, et al. Serum Metabolomics Signatures Associated With Ankylosing Spondylitis and TNF Inhibitor Therapy. *Front Immunol.* 19 févr 2021;12:630791.
28. Kapoor SR, Filer A, Fitzpatrick MA, Fisher BA, Taylor PC, Buckley CD, et al. Metabolic Profiling Predicts Response to Anti-Tumor Necrosis Factor α Therapy in Patients With Rheumatoid Arthritis. *Arthritis Rheum.* juin 2013;65(6):1448-56.
29. Ducourau E, Rispens T, Samain M, Dernis E, Le Guilchard F, Andras L, et al. Methotrexate effect on immunogenicity and long-term maintenance of adalimumab in axial spondyloarthritis: a multicentric randomised trial. *RMD Open.* 9 janv 2020;6(1):e001047.
30. Wolfe F, Hawley DJ. The longterm outcomes of rheumatoid arthritis: Work disability: a prospective 18 year study of 823 patients. *J Rheumatol.* nov 1998;25(11):2108-17.
31. Chorus AMJ. Quality of life and work in patients with rheumatoid arthritis and ankylosing spondylitis of working age. *Ann Rheum Dis.* 1 déc 2003;62(12):1178-84.
32. Dincer U, Cakar E, Kiralp MZ, Bozkanat E, Kilac H, Dursun H. The Pulmonary Involvement in Rheumatic Diseases: Pulmonary Effects of Ankylosing Spondylitis and Its Impact on Functionality and Quality of Life. *Tohoku J Exp Med.* 2007;212(4):423-30.
33. Varan Ö, Babaoğlu H, Göker B. Associations between Depressive Disorders and Inflammatory Rheumatic Diseases. *Curr Top Med Chem.* 2018;18(16):1395-401.

34. Pelechas E, Voulgari PV, Drosos AA. Preclinical discovery and development of adalimumab for the treatment of rheumatoid arthritis. *Expert Opin Drug Discov.* mars 2021;16(3):227-34.
35. van de Putte LBA. Efficacy and safety of adalimumab as monotherapy in patients with rheumatoid arthritis for whom previous disease modifying antirheumatic drug treatment has failed. *Ann Rheum Dis.* 1 mai 2004;63(5):508-16.
36. van der Heijde D, Kivitz A, Schiff MH, Sieper J, Dijkmans BAC, Braun J, et al. Efficacy and safety of adalimumab in patients with ankylosing spondylitis: Results of a multicenter, randomized, double-blind, placebo-controlled trial. *Arthritis Rheum.* juill 2006;54(7):2136-46.
37. Poddubnyy D, Rudwaleit M. Efficacy and safety of adalimumab treatment in patients with rheumatoid arthritis, ankylosing spondylitis and psoriatic arthritis. *Expert Opin Drug Saf.* juill 2011;10(4):655-73.
38. Hyrich KL, Watson KD, Silman AJ, Symmons DPM, British Society for Rheumatology Biologics Register. Predictors of response to anti-TNF-alpha therapy among patients with rheumatoid arthritis: results from the British Society for Rheumatology Biologics Register. *Rheumatol Oxf Engl.* déc 2006;45(12):1558-65.
39. Söderlin MK, Petersson IF, Geborek P. The effect of smoking on response and drug survival in rheumatoid arthritis patients treated with their first anti-TNF drug. *Scand J Rheumatol.* févr 2012;41(1):1-9.
40. on behalf of the Rheumatologists of the Swiss Clinical Quality Management Program, Micheroli R, Hebeisen M, Wildi LM, Exer P, Tamborrini G, et al. Impact of obesity on the response to tumor necrosis factor inhibitors in axial spondyloarthritis. *Arthritis Res Ther.* déc 2017;19(1):164.
41. Law-Wan J, Sparfel MA, Derolez S, Azzopardi N, Goupille P, Detert J, et al. Predictors of response to TNF inhibitors in rheumatoid arthritis: an individual patient data pooled analysis of randomised controlled trials. *RMD Open.* nov 2021;7(3):e001882.
42. Rizzo C, Camarda F, Donzella D, La Barbera L, Guggino G. Metabolomics: An Emerging Approach to Understand Pathogenesis and to Assess Diagnosis and Response to Treatment in Spondyloarthritis. *Cells.* 4 févr 2022;11(3):549.
43. Souto-Carneiro M, Tóth L, Behnisch R, Urbach K, Klika KD, Carvalho RA, et al. Differences in the serum metabolome and lipidome identify potential biomarkers for seronegative rheumatoid arthritis versus psoriatic arthritis. *Ann Rheum Dis.* avr 2020;79(4):499-506.
44. Wang N, Yang L, Shang L, Liang Z, Wang Y, Feng M, et al. Altered Fecal Metabolomics and Potential Biomarkers of Psoriatic Arthritis Differing From Rheumatoid Arthritis. *Front Immunol.* 28 févr 2022;13:812996.
45. Ding NS, McDonald JAK, Perdones-Montero A, Rees DN, Adegbola SO, Misra R, et al. Metabolomics and the Gut Microbiome Associated With Primary Response to Anti-TNF Therapy in Crohn's Disease. *J Crohns Colitis.* 7 sept 2020;14(8):1090-102.
46. Gupta L, Guleria A, Rawat A, Kumar D, Aggarwal A. NMR-based clinical metabolomics revealed distinctive serum metabolic profiles in patients with spondyloarthritis. *Magn Reson Chem.* févr 2021;59(2):85-98.
47. Edwards LJ, Constantinescu CS. Platelet activating factor/platelet activating factor receptor pathway as a potential therapeutic target in autoimmune diseases. *Inflamm Allergy Drug Targets.* juill 2009;8(3):182-90.
48. Hilliquin P, Menkes CJ, Laoussadi S, Benveniste J, Arnoux B. Presence of paf-acether in rheumatic diseases. *Ann Rheum Dis.* 1 janv 1992;51(1):29-31.

49. Camussi G, Bussolino F, Salvidio G, Baglioni C. Tumor necrosis factor/cachectin stimulates peritoneal macrophages, polymorphonuclear neutrophils, and vascular endothelial cells to synthesize and release platelet-activating factor. *J Exp Med.* 1 nov 1987;166(5):1390-404.
50. Bussolino F, Camussi G, Baglioni C. Synthesis and release of platelet-activating factor by human vascular endothelial cells treated with tumor necrosis factor or interleukin 1 alpha. *J Biol Chem.* août 1988;263(24):11856-61.
51. Bonavida B, Mencia-Huerta JM, Braquet P. Effects of platelet-activating factor on peripheral blood monocytes: induction and priming for TNF secretion. *J Lipid Mediat.* 1990;2 Suppl:S65-76.
52. Herrero-Beaumont G, Egido J. PAF, a potent proinflammatory mediator, looking for its role in the pathogenesis of joint damage. *Ann Rheum Dis.* 1 avr 1997;56(4):211-3.
53. Zarco P, Maestre C, Herrero-Beaumont G, González E, Garcia-Hoyo R, Navarro FJ, et al. Involvement of platelet-activating factor and tumour necrosis factor in the pathogenesis of joint inflammation in rabbits. *Clin Exp Immunol.* 28 juin 2008;88(2):318-23.
54. Yu D, Du J, Pu X, Zheng L, Chen S, Wang N, et al. The Gut Microbiome and Metabolites Are Altered and Interrelated in Patients With Rheumatoid Arthritis. *Front Cell Infect Microbiol.* 25 janv 2022;11:763507.
55. O'Shea FD, Tsui FWL, Chiu B, Tsui HW, Yazdanpanah M, Inman RD. Retinol (Vitamin A) and Retinol-Binding Protein Levels Are Decreased in Ankylosing Spondylitis: Clinical and Genetic Analysis. :3.
56. Bidad K, Salehi E, Jamshidi A, Saboor-Yaraghi AA, Oraei M, Meysamie A, et al. Effect of All-transretinoic Acid on Th17 and T Regulatory Cell Subsets in Patients with Ankylosing Spondylitis. *J Rheumatol.* avr 2013;40(4):476-83.
57. Wu J, Wu X, Chen Z, Lv Q, Yang M, Zheng X, et al. Circulating Retinol-Binding Protein 4 as a Possible Biomarker of Treatment Response for Ankylosing Spondylitis: An Array-Based Comparative Study. *Front Pharmacol.* 10 mars 2020;11:231.
58. Young SP, Kapoor SR, Viant MR, Byrne JJ, Filer A, Buckley CD, et al. The Impact of Inflammation on Metabolomic Profiles in Patients With Arthritis: Inflammation and Metabolomic Profile in Arthritis. *Arthritis Rheum.* août 2013;65(8):2015-23.
59. Priori R, Casadei L, Valerio M, Scrivo R, Valesini G, Manetti C. 1H-NMR-Based Metabolomic Study for Identifying Serum Profiles Associated with the Response to Etanercept in Patients with Rheumatoid Arthritis. Driscoll PC, éditeur. *PLOS ONE.* 11 nov 2015;10(11):e0138537.
60. Wang Q, Xu R. MetabolitePredict: A de novo human metabolomics prediction system and its applications in rheumatoid arthritis. *J Biomed Inform.* juill 2017;71:222-8.
61. Zhou J, Chen J, Hu C, Xie Z, Li H, Wei S, et al. Exploration of the serum metabolite signature in patients with rheumatoid arthritis using gas chromatography–mass spectrometry. *J Pharm Biomed Anal.* 5 août 2016;127:60-7.
62. Sasaki C, Hiraishi T, Oku T, Okuma K, Suzumura K, Hashimoto M, et al. Metabolomic approach to the exploration of biomarkers associated with disease activity in rheumatoid arthritis. Mudiam MKR, éditeur. *PLOS ONE.* 11 juill 2019;14(7):e0219400.
63. Takahashi S, Saegusa J, Onishi A, Morinobu A. Biomarkers identified by serum metabolomic analysis to predict biologic treatment response in rheumatoid arthritis patients. *Rheumatology.* 1 déc 2019;58(12):2153-61.
64. Smolenska Z, Smolenski RT, Zdrojewski Z. Plasma concentrations of amino acid and nicotinamide metabolites in rheumatoid arthritis--potential biomarkers of disease activity and drug treatment. *Biomark Biochem Indic Expo Response Susceptibility Chem.* 2016;21(3):218-24.

65. Dimitroulas T, Sandoo A, Kitas GD. Asymmetric Dimethylarginine as a Surrogate Marker of Endothelial Dysfunction and Cardiovascular Risk in Patients with Systemic Rheumatic Diseases. *Int J Mol Sci.* 26 sept 2012;13(10):12315-35.
66. Chandrasekharan UM, Wang Z, Wu Y, Wilson Tang WH, Hazen SL, Wang S, et al. Elevated levels of plasma symmetric dimethylarginine and increased arginase activity as potential indicators of cardiovascular comorbidity in rheumatoid arthritis. *Arthritis Res Ther.* déc 2018;20(1):123.
67. Onmaz DE, Isik K, Sivrikaya A, Abusoglu S, Gezer İA, Abusoglu G, et al. Determination of serum methylarginine levels by tandem mass spectrometric method in patients with ankylosing spondylitis. *Amino Acids.* sept 2021;53(9):1329-38.
68. Kamleh MA, Snowden SG, Grapov D, Blackburn GJ, Watson DG, Xu N, et al. LC-MS Metabolomics of Psoriasis Patients Reveals Disease Severity-Dependent Increases in Circulating Amino Acids That Are Ameliorated by Anti-TNF α Treatment. *J Proteome Res.* 2 janv 2015;14(1):557-66.
69. Takahashi S, Saegusa J, Sendo S, Okano T, Akashi K, Irino Y, et al. Glutaminase 1 plays a key role in the cell growth of fibroblast-like synoviocytes in rheumatoid arthritis. *Arthritis Res Ther.* déc 2017;19(1):76.
70. Gao P, Lu C, Zhang F, Sang P, Yang D, Li X, et al. Integrated GC-MS and LC-MS plasma metabonomics analysis of ankylosing spondylitis. *The Analyst.* 2008;133(9):1214.
71. Berlinberg AJ, Regner EH, Stahly A, Brar A, Reisz JA, Gerich ME, et al. Multi ‘Omics Analysis of Intestinal Tissue in Ankylosing Spondylitis Identifies Alterations in the Tryptophan Metabolism Pathway. *Front Immunol.* 3 mars 2021;12:587119.
72. Smallwood MJ, Nissim A, Knight AR, Whiteman M, Haigh R, Winyard PG. Oxidative stress in autoimmune rheumatic diseases. *Free Radic Biol Med.* sept 2018;125:3-14.
73. Quiñonez-Flores CM, González-Chávez SA, Del Río Nájera D, Pacheco-Tena C. Oxidative Stress Relevance in the Pathogenesis of the Rheumatoid Arthritis: A Systematic Review. *BioMed Res Int.* 31 mai 2016;2016:e6097417.
74. Yang XY, Zheng KD, Lin K, Zheng G, Zou H, Wang JM, et al. Energy Metabolism Disorder as a Contributing Factor of Rheumatoid Arthritis: A Comparative Proteomic and Metabolomic Study. *PLoS ONE.* 6 juill 2015;10(7):e0132695.
75. Ye G, Xie Z, Zeng H, Wang P, Li J, Zheng G, et al. Oxidative stress-mediated mitochondrial dysfunction facilitates mesenchymal stem cell senescence in ankylosing spondylitis. *Cell Death Dis.* 17 sept 2020;11(9):775.
76. Wang L, Gao L, Jin D, Wang P, Yang B, Deng W, et al. The Relationship of Bone Mineral Density to Oxidant/Antioxidant Status and Inflammatory and Bone Turnover Markers in a Multicenter Cross-Sectional Study of Young Men with Ankylosing Spondylitis. *Calcif Tissue Int.* juill 2015;97(1):12-22.
77. Wahl SM, McCartney-Francis N, Chan J, Dionne R, Ta L, Orenstein JM. Nitric Oxide in Experimental Joint Inflammation. *Cells Tissues Organs.* 2003;174(1-2):26-33.
78. Yamamoto K, Chiba T, Matsumoto T. Effect of tumor necrosis factor- α antagonists on oxidative stress in patients with Crohn’s disease. *World J Gastroenterol.* 21 sept 2015;21(35):10208-14.
79. Psychogios N, Hau DD, Peng J, Guo AC, Mandal R, Bouatra S, et al. The Human Serum Metabolome. *PLoS ONE.* 16 févr 2011;6(2):e16957.
80. England BR, Thiele GM, Anderson DR, Mikuls TR. Increased cardiovascular risk in rheumatoid arthritis: mechanisms and implications. *The BMJ.* 23 avr 2018;361:k1036.

81. Wang R, Ward MM. EPIDEMIOLOGY OF AXIAL SPONDYLOARTHRITIS: AN UPDATE. *Curr Opin Rheumatol.* mars 2018;30(2):137-43.
82. Mittelstrass K, Ried JS, Yu Z, Krumsiek J, Gieger C, Prehn C, et al. Discovery of Sexual Dimorphisms in Metabolic and Genetic Biomarkers. McCarthy MI, éditeur. PLoS Genet. 11 août 2011;7(8):e1002215.
83. He Z, Wang M, Li H, Wen C. GC-MS-based fecal metabolomics reveals gender-attributed fecal signatures in ankylosing spondylitis. *Sci Rep.* 7 mars 2019;9(1):3872.
84. Fu J, Cuppen BJ, Welsing PMJ, van Wietmarschen H, Harms AC, Berger R, et al. Differences between serum polar lipid profiles of male and female rheumatoid arthritis patients in response to glucocorticoid treatment. *Inflammopharmacology.* déc 2016;24(6):397-402.

Annexes

- I. Diagnostic criteria for Spondylarthritis by Assessment of Spondyloarthritis international Society (ASAS) and diagnostic criteria for Rheumatoid Arthritis by American College of Rheumatology (ACR) 1987

1. Diagnostic criteria for SpA:

The diagnosis can be made at onset **in subjects < 45 years of age with back pain for at least 3 months** if the following are respected:

- **Sacroiliitis on imaging* + ≥ 1 SpA feature OR HLA-B27 + ≥ 2 other SpA features**

SpA features: inflammatory back pain; arthritis; enthesitis (heel); uveitis; dactylitis; psoriasis; Crohn's/colitis; good response to non-steroid anti-inflammatory drugs (NSAIDs); family history for SpA; HLA-B27; elevated CRP

*: sacroiliitis on imaging means: active (acute) inflammation on MRI highly suggestive of sacroiliitis associated with SpA OR definite radiographic sacroiliitis according to the modified New York criteria.

2. Diagnostic criteria for Rheumatoid Arthritis by ACR 1987

A patient is considered to have rheumatoid arthritis if he meets 4 of the 7 criteria.

Table 1. ACR 1987 diagnostic criteria for Rheumatoid Arthritis.

1. Morning stiffness	Morning stiffness in and around the joint lasting at least one hour before maximal improvement, persisting > 6 weeks.
2. Arthritis of 3 or more joint areas	At least three joints' areas simultaneously have had soft tissue swelling for fluid observed by a physician, persisting > 6 weeks.
3. Arthritis of hand joints	At least one swollen joint in a wrist, MCP joint or IPP joint persisting > 6 weeks.
4. Symmetrical arthritis	Simultaneous involvement of the same joint area on both sides of the body persisting > 6 weeks.
5. Rheumatoid nodules	Subcutaneous nodules, over bony prominences, or extensor surfaces or in juxta-articular regions, observed by a physician.
6. Serum rheumatoid factor	Positive serum RF
7. Radiographic changes	Radiographic changes typical of rheumatoid arthritis on posteroanterior hand and wrist radiographs, which must include erosions and bony decalcification

- II. Spondylarthritis (ASDAS) and rheumatoid arthritis (DAS28) activity score with assessment of disease activity according to this score.

The ASDAS score is a composite score calculated with the following formula:

$$\text{ASDAS-CRP} = 0.12 \times \text{spinal pain} + 0.06 \times \text{duration spinal stiffness} + 0.11 \times \text{overall patient assessment} + 0.07 \times \text{peripheral pain/swelling} + 0.58 \times \ln(\text{CRP}+1).$$

Depending on the result obtained, one could speak of:

- ASDAS-CRP < 1.3: inactive disease
- 1.3 < ASDAS-CRP < 2.1: disease of moderate activity
- 2.1 < ASDAS-CRP < 3.5 : active disease
- ASDAS-CRP > 3.5 : very active disease

The DAS28 score is also a composite score considering the articular index (painful joints) and the synovitis index (swollen joints) of the 28 joints of the EULAR, the sedimentation rate, the evaluation of the overall condition of the patient on a visual analogue scale. It is calculated according to the following formula:

$$\text{DAS28} = 0.55 \times \sqrt{(\text{tender joint count } 28)} + 0.284 \times \sqrt{(\text{swollen joint count } 28)} + 0.37 \log (\text{ESR}) + 0.0142 \times (\text{General health}).$$

Depending on the result obtained, the disease was defined as:

- DAS28 < 2.6 : remission
- 2.6 < DAS28 < 3.2 : low activity
- 3.2 < DAS28 < 5.1 : moderate activity
- DAS28 > 5.1 : high activity

III. Classification of patients into two groups according to ASDAS response and DAS28 response (EULAR criteria).

ASDAS at each visit	Improvement of ASDAS compared to W0		
	< 1.1	≥ 1.1 and < 2	≥ 2
< 1.3		Good responder	Very good responder
≥ 1.3 et < 2.1		Moderate responder	Good responder
≥ 2.1 et < 3.5	No responder	Moderate responder	Moderate responder
≥ 3.5	No responder	No responder	No responder

Table 2. Classification of patients according to the change in ASDAS score at each visit compared to W0.

DAS28 at each visit	Improvement of DAS28 compared to W0		
	> 1.2	> 0.6 and < 1.2	≤ 0.6
≤ 3.2	Good responder		
> 3.2 and ≤ 5.1		Moderate responder	
> 5.1			Non-responder

Table 3. Classification of patients according to the change in DAS28 at each visit compared to W0 by EULAR criteria.

IV. Outliers at different analysis times

COMARIS

Analysis of the metabolome at W0 according to the therapeutic response at W12:

- PCA: a woman, responder, treated with adalimumab + methotrexate; a woman, non-responder, treated with adalimumab alone.
- PLS-DA: both PCA patients + one man, responder, treated with adalimumab alone.

Analysis of the metabolome at W0 according to the therapeutic response to W26:

- PCA: one man, responder, treated with adalimumab alone; three women including 2 responders, treated with adalimumab + methotrexate and one, non-responder treated with adalimumab alone.
- PLS-DA: the man responder of the PCA; a female responder, treated with adalimumab + methotrexate; a non-responding woman, treated with adalimumab alone (patients found in PCA).

Analysis of the variation of the metabolome between W0 and W4 according to the therapeutic response to W12:

- PCA : none.
- PLS-DA: a woman, non-responders, treated with adalimumab alone; a female responder, treated with adalimumab + methotrexate.

Analysis of the variation of the metabolome between W0 and W4 according to the therapeutic response to W26:

- PCA: a woman, non-responder, treated with adalimumab alone.
- PLS-DA: a woman, responder, treated with adalimumab + methotrexate; a man responder, treated by adalimumab alone.

AFORA

Analysis of the W0 metabolome according to the therapeutic response to W12:

- PCA: 4 women, 3 responders treated with either adalimumab alone, adalimumab + methotrexate or adalimumab + corticosteroids and one non-responder treated with adalimumab + methotrexate + corticosteroids.
- PLS-DA: 3 women, 2 responders treated with adalimumab alone and one non-responder treated with adalimumab + methotrexate + corticosteroids.

Analysis of the W0 metabolome according to the therapeutic response to W26:

- PCA: 4 women, responders, treated with adalimumab alone, adalimumab + methotrexate, adalimumab + corticosteroids and adalimumab + methotrexate + corticosteroids.
- PLS-DA: two women including one responder and one non-responder, treated with adalimumab + corticosteroids; one man, responder, treated with adalimumab + methotrexate + corticosteroids.

Analysis of the variation of the metabolome between W0 and W4 according to the therapeutic response to W12:

- PCA: 2 women, responders, treated with adalimumab + methotrexate and adalimumab alone respectively.
- PLS-DA: 4 women, responders including 3 treated with adalimumab + methotrexate and the last with adalimumab alone.

Analysis of the variation of the metabolome between W0 and W4 according to the therapeutic response to W26:

- PCA: a woman, responder, treated with adalimumab + methotrexate
- PLS-DA: 2 women, responders including one treated with adalimumab + methotrexate (same as PCA) and the other treated with adalimumab alone.

V. Figures

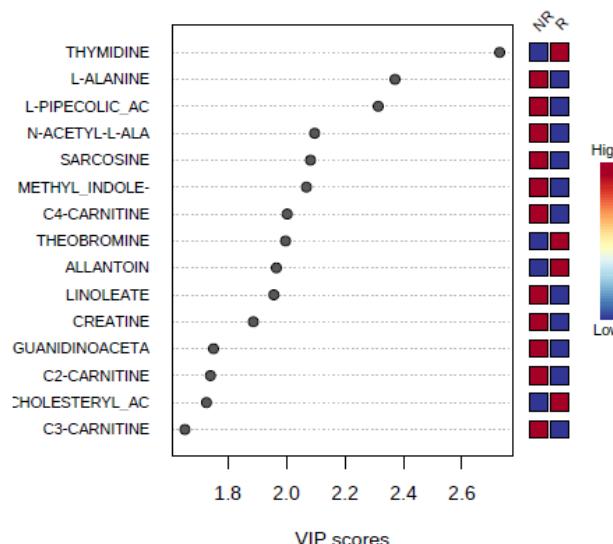


Figure 1. COMARIS. VIP-scores in PLS-DA analysis of the metabolic profile at W0 according to the therapeutic response at 12 weeks. R = responders; NR = non-responders.

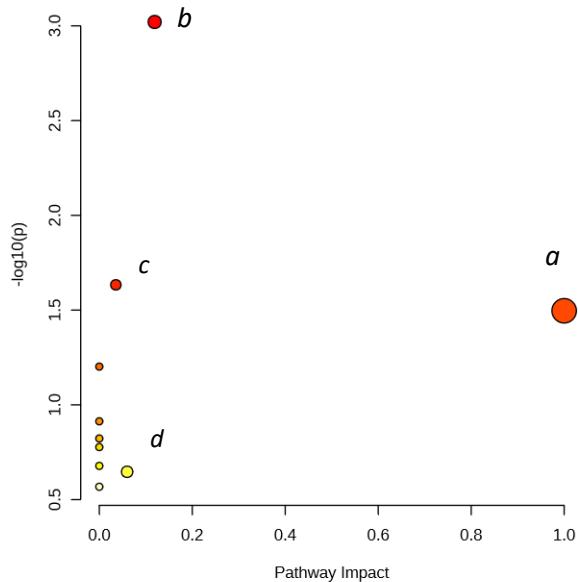


Figure 2. COMARIS. Impact of W0 metabolic pathways between responder and non-responder patients at 12 weeks. a: linoleic acid metabolism; b: glycine, serine, and threonine metabolism; c: arginine and proline metabolism; d: pyrimidine metabolism.

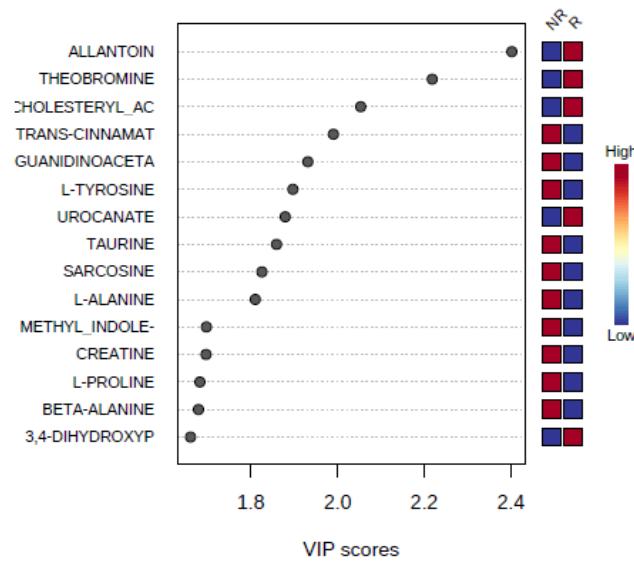


Figure 3. COMARIS. VIP-scores in PLS-DA analysis of the metabolic profile at W0 according to the therapeutic response at 26 weeks. R = responders; NR = non-responders.

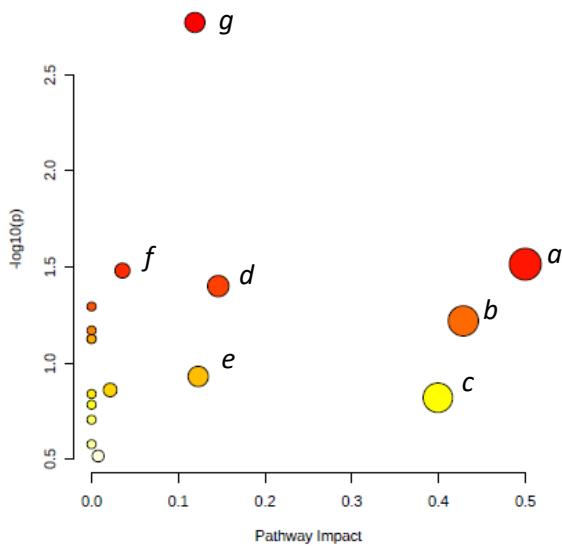


Figure 4. COMARIS. Impact of metabolic pathways involved in W0 between responder and non-responder patients at 26 weeks. a: phenylalanine, tyrosine and tryptophan biosynthesis; b: taurine and hypotaurine metabolism; c: beta-alanine metabolism; d : tyrosine metabolism ; e : histidine metabolism ; f : arginine and proline metabolism ; g : glycine, serine and threonine metabolism.

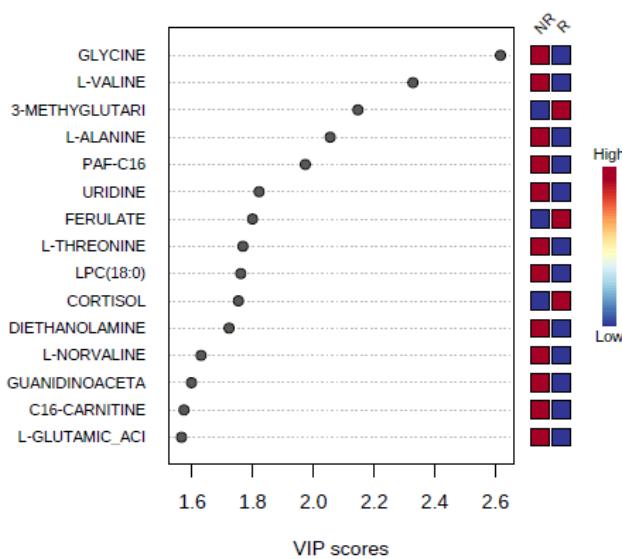


Figure 5. COMARIS. VIP-scores in PLS-DA analysis of the change in metabolome between W0 and W4 according to therapeutic response at 26 weeks. R = responders; NR = non-responders.

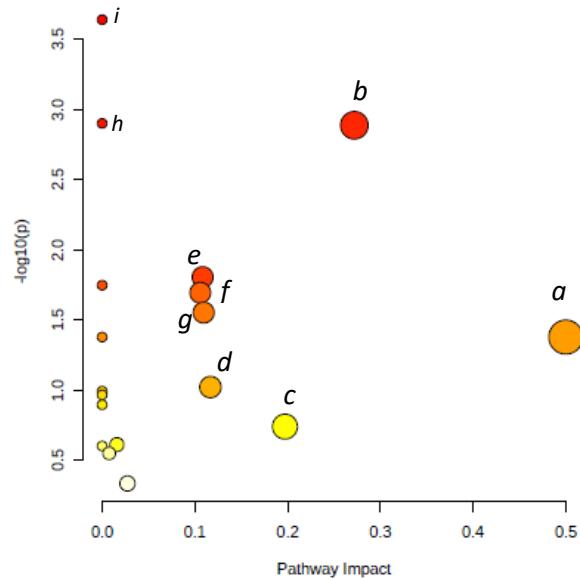


Figure 6. COMARIS. Impact of metabolic pathways involved in metabolome modification between W0 and W4 according to therapeutic response at 26 weeks. a: D-glutamine and D-glutamate metabolism; b: glycine, serine and threonine metabolism; c : alanine, aspartate and glutamate metabolism ; d : arginine biosynthesis ; e : glutathione metabolism ; f : glyoxylate and dicarboxylate metabolism ; g : arginine and proline metabolism ; h : valine, leucine and isoleucine biosynthesis ; i : aminoacyl-t-RNA biosynthesis.

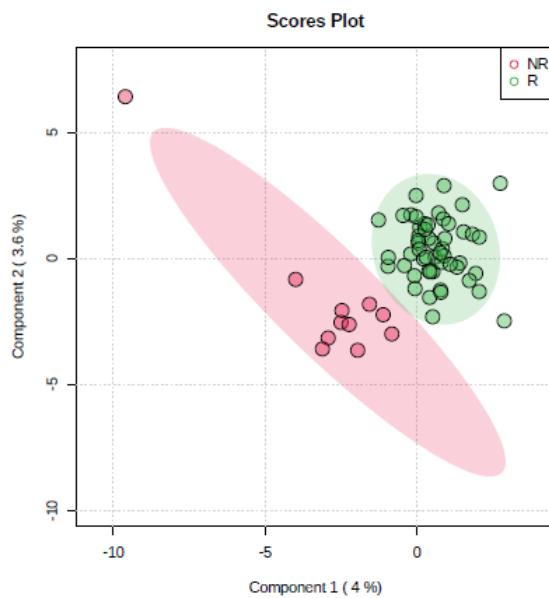


Figure 7. AFORA. Distribution of the W0 metabolome of different patient groups to PLS-DA according to response at 12 weeks. R = responders; NR = non-responders.

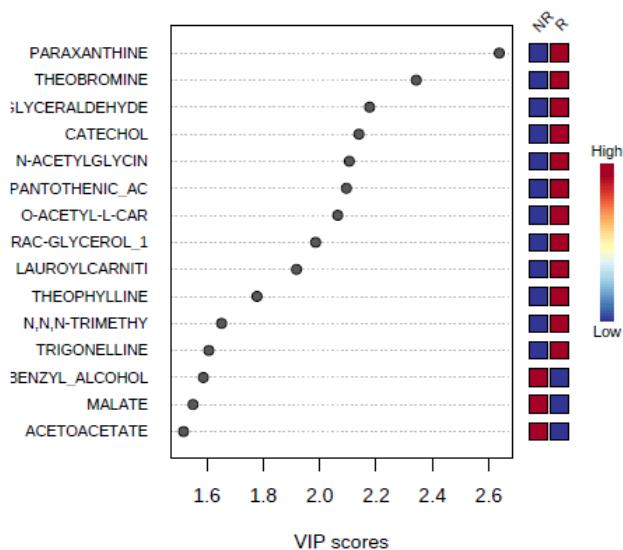


Figure 8. AFORA. VIP-score in PLS-DA analysis of the W0 metabolome in different patient groups according to therapeutic response at 12 weeks. R = responders; NR = non-responders.

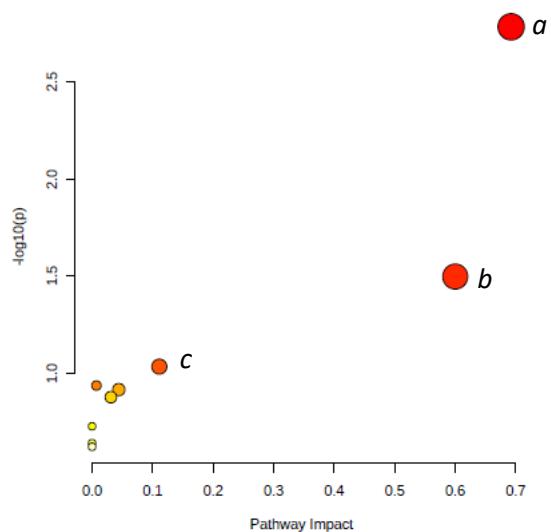


Figure 9. AFORA. Impact of metabolic pathways involved at W0 between responder and non-responder patients at 12 weeks. a: caffeine metabolism; b: synthesis and degradation of ketone bodies; c: butanoate metabolism.

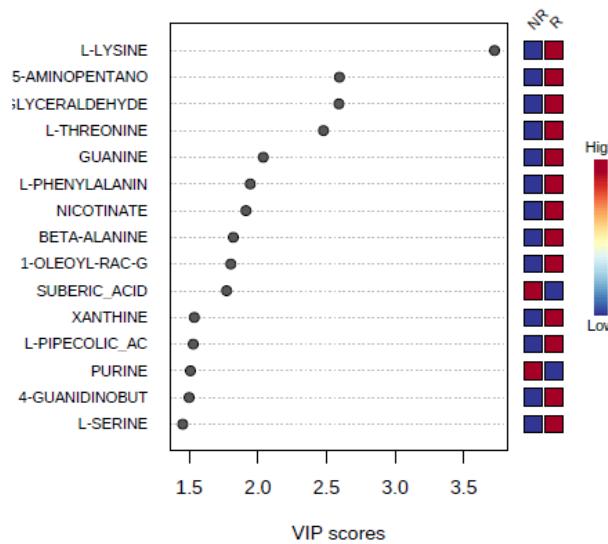


Figure 10. VIP-score in PLS-DA analysis of the W0 metabolome in different patient groups according to therapeutic response at 26 weeks. R = responders; NR = non-responders.

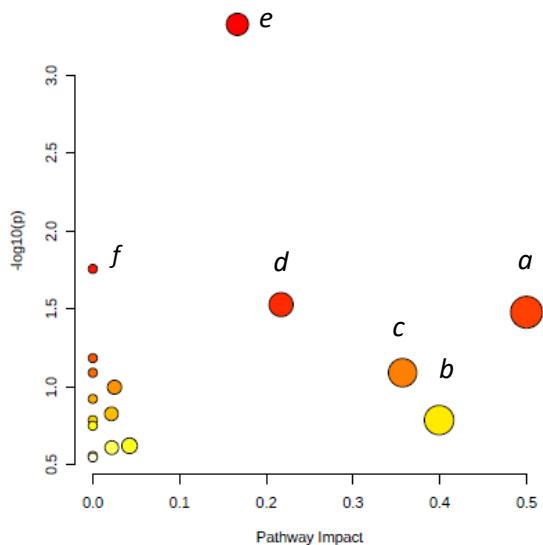


Figure 11. AFORA. Impact of metabolic pathways involved at W0 between responder and non-responder patients according to therapeutic response at 26 weeks. a: phenylalanine, tyrosine, and tryptophan metabolism; b: beta-alanine metabolism; c: phenylalanine metabolism; d: glycine, serine, and threonine metabolism; e: aminoacyl-tRNA biosynthesis; f: lysine degradation.

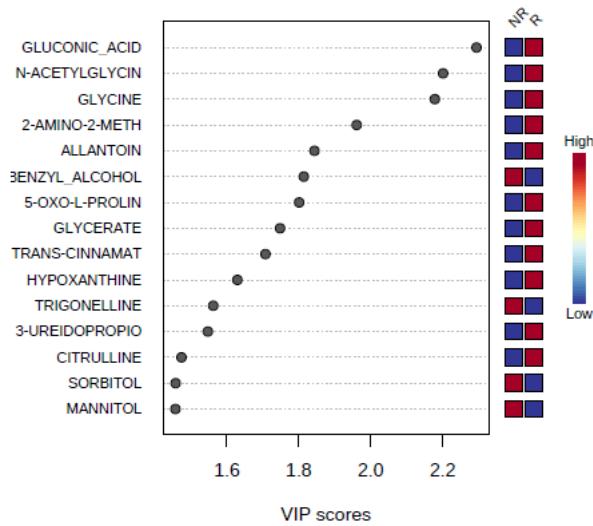


Figure 12. AFORA. VIP-score in PLS-DA analysis of the change in the metabolome between W0 and W4 in different patient groups according to the therapeutic response at 26 weeks. R = responders; NR = non-responders.

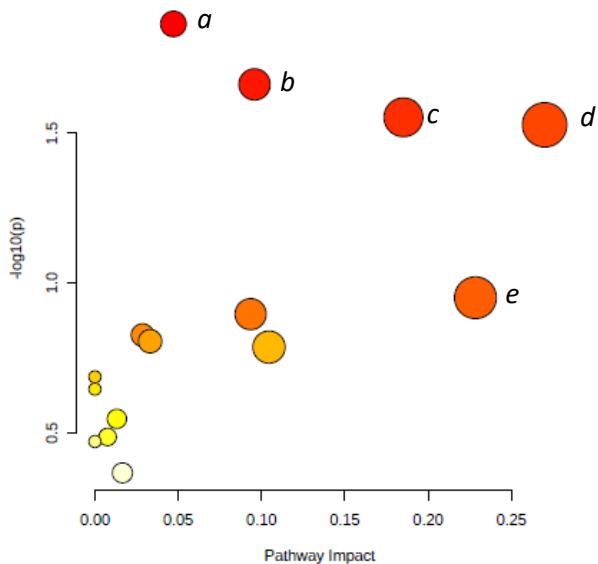


Figure 13. AFORA. Impact of metabolic pathways involved in metabolome modification between W0 and W4 according to therapeutic response at 26 weeks. a: pentose phosphate pathway; b: glutathione metabolism; c: glyoxylate and dicarboxylate metabolism; d: glycine, serine, and threonine metabolism; e: arginine biosynthesis.

Vu, les Directeurs de Thèse



Vu, le Doyen
De la Faculté de Médecine de Tours
Tours, le

Elom Annette Amie TAY

52 pages – 1 tableau – 10 figures – 5 annexes.

RESUME

Objectifs : Analyser les changements métabolomiques 4 semaines après l'initiation de l'adalimumab, un anti-TNF chez les patients atteints de spondylarthrite axiale (SpA) et de polyarthrite rhumatoïde (PR) et étudier la relation avec la réponse clinique à 12 et 26 semaines.

Méthodes : Nous avons effectué des analyses en post-hoc du sérum de patients atteints de SpA de l'étude COMARIS (NCT01895764) et de patients atteints de PR de l'étude AFORA (NCT01382160) par chromatographie liquide couplée à la spectrométrie de masse à haute résolution (LC/HRMS) à l'inclusion (S0) et 4 semaines (S4) après l'initiation du traitement par adalimumab. Les réponses cliniques ont été évaluées à 12 et 26 semaines. Les variations des concentrations de métabolites entre S0 et S4 ont été comparées entre les répondeurs et les non-répondeurs en analyse univariée, multivariée non supervisée en composantes principales (PCA) puis multivariée supervisée par des analyses discriminantes partielles des moindres carrés (PLS-DA), à l'aide du logiciel METABOANALYST. Les variables d'intérêt, définies comme celles ayant un score VIP (Variable Influence on Projection) supérieur ou égal à 2 ont été sélectionnées. Enfin, nous avons étudié les différentes voies métaboliques dans lesquelles ces variables d'intérêt étaient impliquées.

Résultats : Soixante-quatorze patients ont été inclus dans l'étude COMARIS, dont 43 répondeurs et 31 non-répondeurs. Soixante-trois patients ont été inclus dans l'étude AFORA, dont 52 répondeurs et 11 non-répondeurs. En analyse univariée, la leucine, l'hypoxanthine et la n-acétyl-l-alanine étaient abaissées chez les répondeurs atteints de SpA. La carnosine, le cortisol et la purine étaient abaissés chez les répondeurs atteints de PR, tandis que la l-méthionine et la 5-oxo-l-proline étaient augmentées chez ces sujets. Nous n'avons pas été en mesure de différencier les patients de chaque étude en fonction de leur réponse en utilisant la PCA et la PLS-DA. Cependant, chez les patients atteints de SpA, cinq métabolites avec un score VIP >2 de l'analyse PLS-DA, y compris le PAF-C16, un facteur d'activation plaquettaire, et le LysoPC (18:0), un composant des membranes cellulaires, semblaient pertinents. Diverses voies métaboliques ont été identifiées dans les deux études : métabolisme des glycérophospholipides ; métabolisme de l'arginine et de la proline; métabolisme de lalanine, de l'aspartate et du glutamate; biosynthèse de la phénylalanine, de la tyrosine et du tryptophane; métabolisme de la cystéine et de la méthionine; biosynthèse du pantothenate et du CoA et biosynthèse de l'aminoacyl-ARNt.

Conclusion : Chez les patients atteints de PR et de SpA, l'adalimumab induit des changements précoce dans le métabolome impliquant des voies telles que le métabolisme des acides aminés essentiels et non essentiels et le stress oxydatif. Ce résultat pourrait guider les investigations futures afin de trouver des marqueurs prédictifs de la réponse thérapeutique aux anti-TNF dans les rhumatismes inflammatoires chroniques.

Mots-clés : métabolomique ; spondylarthrite axiale ; polyarthrite rhumatoïde ; adalimumab ; prédiction de la réponse ; LC/HRMS ; acides aminés ; stress oxydatif.

Jury :

Président du jury : Professeur Philippe GOUILLLE

Directeurs de thèse : Professeur Denis MULLEMAN & Professeur Hélène BLASCO

Membres du jury : Professeur Jérémie SELLAM

Professeur Patrick EMOND

Date de soutenance : lundi 20 juin 2022.